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BIOLOGICAL BEHAVIOR OF 60/40 BIPHASIC CALCIUM PHOSPHATE AND GROWTH HORMONE MIXTURE: IN VIVO STUDY IN DOGS

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ABSTRACT

Grafting materials are important for ridge preservation procedures after tooth extraction. Ridge preservation prime value is the volume preservation for the healing sockets, which is needed for different ridge augmentation procedures.

Objective: Determine the healing responses of (local growth hormone, and biphasic calcium phosphate mixture 60/40 in fresh extraction sockets of a canine model.

Materials and Methods: Six mongrel dogs included in this study; they were divided into 2 groups according to the time of sacrifice (4 and 8 weeks). Each of these groups includes three animals. Animals subjected to extraction of the fourth premolar bilaterally, The right side treated with a mixture of 4 IU (1.6 mg) Growth hormone and biphasic calcium phosphate (a composite of hydroxyapatite HA and bèta-tricalcium phosphate TCP 1 x 0.15 ml (450-650 μ m) in a granular form, while the left side left as a control. The healing extraction sockets were evaluated by surface electron microscope (SEM).

Results: Scanning electron microscope examination showed homogenous new bone formation in both groups. Control group showed significant results as regard calcium phosphate ratio in the first month, while non-significant result was encountered in the second month between two groups.

Conclusion: Single local dose of growth hormone mixed with 60/40 biphasic calcium phosphate did not improve calcium phosphate ratio.

KEYWORDS: Calcium phosphate ratio, surface electron microscope, ridge preservation, biphasic calcium phosphate and dogs.

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INTRODUCTION

Alveolar bone remodeling is a tooth-dependent process. Once the tooth extracted, trauma stimulates the activity of osteoclasts leading to alveolar bone resorption. Resultant clinical bone loss in height and width leads to essential problems for restorative and implant dentistry at the extraction site. The effect is magnified when multiple teeth were extracted in the area. (1-5)

Preservation procedures using various bone substitutes are recommended. Synthetic bone grafts were preferable for their availability, and low risk for disease transmission. Basically, they are made of hydroxyapatite (HA) or tricalcium phosphate (TCP), they have different biodegradability. The rationale for the development of biphasic calcium phosphate bioceramics has been their similarity in composition to bone mineral phase and biodegradability, bioactivity and osteoconductivity. Moreover, biphasic calcium phosphate bioceramics (60 HA/40 beta-TCP) has the advantage of adequate mechanical strength and reasonable resorption rate that allow new bone deposition (6-15)

Growth hormone (GH) local administration in a single dose at time of surgery is promising. It could regulate local bone metabolism by enhancing the proliferation of osteoblastic cell lines, and express bone morphogenetic proteins (BMP) especially in the early stages of healing. (2, 16-18)

Modern computers availability had enabled the scanning electron microscope (SEM), which is important for the observation of surface topography. Moreover, it determines the chemical composition of a specimen by using energy dispersive x-ray analysis (EDX), having the advantages of minimum specimen preparation, relatively quick and nondestructive mean (19,20).

This study used a combination of biphasic calcium phosphate 60/40 and growth hormone at the fresh extraction site for the purpose of early new bone formation and volume preservation.

MATERIALS AND METHODS

The experimental study was performed at the Department of Surgery, Anesthesiology, and Radiology, Faculty of Veterinary Medicine, Suez Canal University. Six adult healthy male mongrel dogs with an average weight of 10-20 kg, comparable age (about 1year) included in this study. Animals divided into 2 groups according to the time of sacrifice (4 and 8 weeks). Each of these groups includes three animals.

All surgical procedures performed using injectable general anesthesia, each dog premedicated with intramuscular injection of Valpam (*Amoun Pharmaceutical Industries co. (APIC) Cairo, Egypt.*) in a dose of 0.5mg / Kg body weight 10-15 min. before operation. Induction and maintenance anesthesia carried out using IV administration of Sodium thiopental (*Egyptian International Pharmaceutical Industries co(EIPICO) 10* th of Ramadan city, EGYPT) 2.5% solution 20-30 mg/kg body weight.

Injection of prophylactic antibiotic carried out using Flumox (*Egyptian International Pharmaceutical Industries co(EIPICO) 10* th of Ramadan city ,EGYPT.) 500mg, 2ml IV every 24 hours for 5 days

Animals subjected to extraction of the fourth premolar bilaterally (Fig.1A&B). The right side treated with a mixture of 4 IU (1.6 mg) Growth hormone (Somatropin 4 IU Sedico Pharmaceutical Industries co,6 th October city.EGYPT) and easygraft- crystal (Wagistrasse 23 - 8952 Schlieren/ Zurich-Switzerland) 1 x 0.15 ml (450-650 μ m) in a granular form, while the left side left as a control (Fig.2A). Releasing incisions followed for primary wound closure using 4.0 vicryl sutures (Fig.2B). Postoperative analgesic (CATAFLAM ®NOVARTIS PHARMA Pharmaceutical industries co,CAIRO,EGYPT.) 75mg / 2ml IM administrated once a day for 3 days. Plaque control with chlorohexidine (ANTISEPTOL KAHIRA, Cairo, Egypt) mouth lavage was used.



Fig. (1) A: Showing 4th premolar tooth. B: Showing the extraction socket.



Fig. (2) A: Showing biphasic calcium phosphate granules in the socket. B: Showing primary closure.

Animal sacrifice performed by rapid IV injection of thiopental sodium overdose. The mandibles dissected, sectioned and put in neutral formalin 10%.

Histomorphometric analysis

1-SEM analysis

Bone blocks were removed from formalin and gently washed before analysis under distilled water, then accurately dehydrated. The studied samples were undersized (1½ x1 cm) to match the sensors inside the device (**Fig.3 A**). The assessment done for the new bone formed midway in the socket (**Fig.3 B**) and the bone proper around socket for each sample.

The scanning voltage was 80 Kv, and the exposure time was 1 second. A total of 1600 projections were acquired, while the specimen was

rotated about 360°. The pixel size was 22.4814 μ m. Then, the visualization and quantification of the newly formed bone were fulfilled on Avizo Fire (Visualization Sciences Group, FEI Corp., OR) image analysis software.

Statistical analysis

Statistical analysis was performed with IBM® SPSS® (SPSS Inc., IBM Corporation, NY, and USA) Statistics Version 24 for Windows. Data presented as Mean, standard deviation (SD. Data explored for normality using Shapiro-Wilk tests. Data for chemical analysis showed nonparametric distribution, so Mann-Whitney test used to compare different groups. Wilcoxon Signed rank test used to compare between different follow-up periods. The significance level was set at $P \le 0.05$.

(182) E.D.J. Vol. 65, No. 1 *Lama H. El Marsafy, et al.*

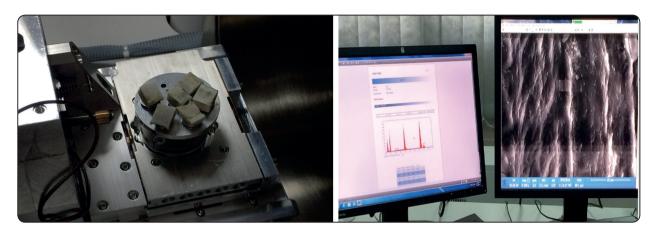


Fig. (3) A: Specimen inside the SEM sensor. B; Software analysis for each specimen

RESULTS:

1. Scanning Electron Microscope Examination at one month and 2 months respectively:

The scanning electron microscope examination showed homogenous intimate contact new bone formation in both groups (**Figures from 4 -7**).

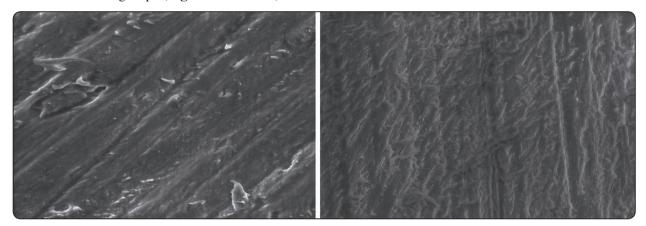


Fig. (4) Scanning electron microscope showed: (a) Control bone surface at 1 month (b) bone surface treated with (HA/TCP) -GH composite at 1 month (X 50)

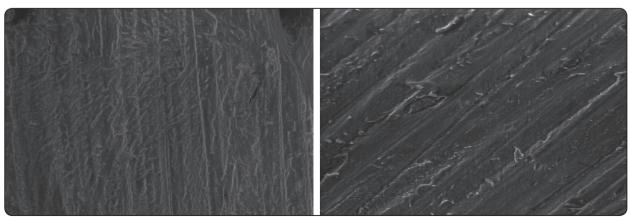


Fig. (5) Scanning electron microscope showed: (a) Control bone surface at 1 month (b) bone surface treated with (HA/TCP) -GH composite at 1 month(X 100)

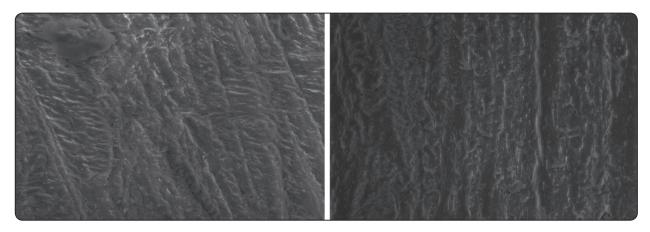


Fig. 6: Scanning electron microscope showed: (a) Control bone surface at 2 month (b) bone surface treated with (HA/TCP) -GH composite at 2 month (X 50)

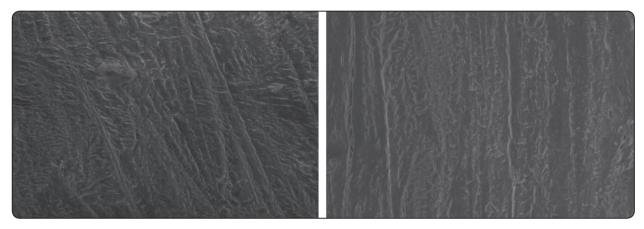


Fig. 7: Scanning electron microscope showed: (a) Control bone surface at 2 month (b) bone surface treated with (HA/TCP) -GH composite at 2 month (X 100)

2. Energy Dispersive Spectrometry

The energy dispersive spectrometry (EDS) analysis of the HA/TCP and GH composite showed chemical analysis composition in form of calcium (CA)/phosphate (P) ratio (**Fig. 8**), and **Tables 1**

I- Comparison between Ca/P ratio in control and test groups

At one month, a higher mean value was recorded in control group, with a significant difference (P=0.017), (**Table 1, Fig. 9**)

At two months, a higher mean value was

recorded in test group, with no significant difference (P=0.78), (**Table 1, Fig. 9**)

II-Comparison between Ca/P ratio in bone proper and both groups

At one month, a higher mean value was recorded in bone proper, with an extremely significant difference (P<0.0001), (Table 2, Fig. 9)

At two months, a slightly higher mean value was recorded in test in comparison to original bone, with no significant difference (P=0.86, P=0.35 for control and test group respectively), (Table 2, Fig. 9)

TABLE (1): Comparison between Ca/P ratio in control and test groups

Groups	CA/P ratio means	P	
1M Control	2.07±0.02	0.017*	
1M Test	2.01±0.04	0.017*	
2M Control	2.27±0.05	0.70ns	
2M Test	2.28±0.06	0.78 ^{ns}	

Significance level P<0.05,*significant,ns=non-significant

TABLE (2) Comparison between Ca/P ratio in Bone proper, control and test groups (Unpaired t- test)

Groups	CA/P ratio	Bone proper	P
1M Control	2.07±0.02	2.3±0.05	<0.0001*
1M Test	2.01 ±0.04		<0.0001*
2M Control	2.27±0.05	2.22±0.12	0.86 ^{ns}
2M Test	2.28±0.06		0.35 ^{ns}

Significance level P<0.05,*significant,ns=non-significant

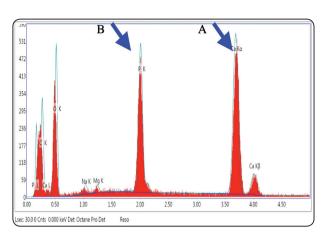


Fig. (8) EDX spectrum showing calcium(CA) and phosphate (P) heights

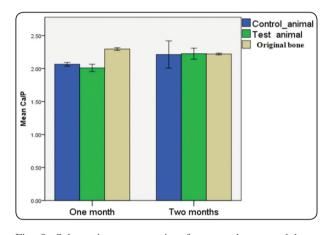


Fig. 9: Schematic representation for control, test and bone proper.

DISCUSSION

Calcium phosphate bioceramics application has been investigated in vitro and in vivo in various studies. It is a biocompatible material having osteoconductive properties which provide suitable environment for bone formation in vivo. (21 - 27)

Biphasic calcium phosphates (BCP) bioceramics granules determine the ideal balance between stable phase HA and resorbable phase β -TCP to control in vivo biodegradability. However, there is always a debate as regard the optimum composition ratio. $^{(28-30)}$

Calcium and phosphate are the major mineral components in bone. Measuring the Ca/P ratio is a good index of bone quality. Relationship had been confirmed between bone loss and lowered CA/P ratio. Changes in CA/P ratio is a good indicator for changes in the organic matrix of bone, specially collagen.

Various BCP composition ratios have been studied. Two ratios of HA/b-TCP (50/50 and 60/40) were under clinical trial in humans. Brodie et al. reported that collagen coating increases the strength of HA/TCP 50/50, whereas it weakens HA/TCP

25/75 and pure TCP discs invitro study. This may be related to increased TCP ratio. While Tanimoto Y et al 2009 proved that the ratio of 25/TCP 75 had the best cellular activity in vitro study. (35,36)

The suggested ratio in this study was 60/40 ratio of (HA/ β -TCP) supposed to provide a balance a regard the strength during growth hormone addition as a source of collagen.

Therefore, development of organic-inorganic hybrid BCP scaffolds provides excellent possibilities for optimizing the conventional bone substitutes. (37) Ying et 2018, evaluated the osteoinductive property of the calcium phosphate bioceramics. He found that the synergistic effect of biphasic calcium phosphate ceramics on seeded macrophages will enhance directed migration of bone marrow-derived mesenchymal stem cells toward the ceramic surface. These stem cells will be stimulated to differentiate into vascular endothelial cells leading to the formation of new blood vessels enhancing new bone. (38)

Biphasic calcium phosphates have been used as a drug delivery systems and tissue engineering scaffolds. (39, 40) They were degraded into CA and phosphate those used for benefit of new bone formation. There crystals exhibit a surface interaction and physicochemical properties making them potential candidates for local delivery of various drugs as hormones, steroids, antibiotics and BMP-2. (41- 44) From all above, this study supposed" GH as a collagen source and as drug to be carried on top of BCP leads to osteoprogenitor cell differentiation with upregulation of osteoblast activity. (45- 47)

Ko-Ning et al 2016 studied the performance of ceramic composite composed of purified fibrillar collagen and HA/TCP in socket preservation in beagle dogs. At eight weeks there scanning electron microscope examination showed that HA/TCP granules were homogenously distributed with the collagen matrix. At the same time, energy dispersive spectrometry analysis of the HA/TCP collagen

composite showed CA/P ratio equal to 1.83. (48) These results were consistent with our study as at two months scanning electron microscope examination showed homogenous new bone formation in both groups (test and control). The energy dispersive spectrometry (EDS) analysis of the HA/TCP/GH composite, showed chemical analysis composition of calcium CA/P ratio equal 2.28±0.06 at test group and 2.27±0.05 at control group. In 2014 Rander et al used histological and scanning electron microscopy (SEM) analysis to evaluate the bone integration of biphasic calcium phosphate BCP bioceramic in rats extraction sockets. There results showed that BCP 60/40 bone graft is a biocompatible material as histological results showed no foreign body reactions or any persistent inflammation. At 7 days, SEM microscopy noticed close contact between connective tissue and bioceramic particles. In 21 to 42 days, scanning electron microscopy analysis showed newly formed bone adhered to ceramic interface with no interposition of soft tissue. (49) This was also obtained by Kitsugi et al 1995 and Fujita et al 2014, who found direct connection between bioceramics and newly formed osteoid matrix. (50,51)

In conclusion, biphasic calcium phosphate and growth hormone (most of its components were collagen) BCP/GH mixture, in one single dose at time of surgery did not make a difference as regard the rate of new bone deposition.

REFERENCES

- Devlin H, Ferguson MJ. Alveolar ridge resorption and mandibular atrophy. A review of the role of local and systemic factor. Br Dent J 1991;170:101-104.
- Lekovic V, Kenney EB, Weinlaender M, Han T, Klokkevold P, Nedic M, Orsini M. A bone regenerative approach to alveolar ridge maintenance following tooth extraction. Report of 10 cases. J Periodontol. 1997;68(6):563–70.
- Utari K., Maretaningtias D., Eha D., Nike H.. The potential of mangosteen (Garcinia mangostana) peel extract, combined with demineralized freeze dried bovine bone xenograft, to reduce ridge resorption and alveolar

- bone regeneration in preserving the tooth extraction socket. Department of Prosthodontics, Faculty of Dental Medicine, Airlangga University, Surabaya, Indonesia. The Journal of Indian Prosthodontic Society | Volume 17 | Issue 3 | July-September 2017.
- 4. Utari K., Tika R., Rostiny R.. Effects of mangosteen peel extract combined with demineralized freeze dried bovine bone xenograft on osteocalcin, collagen 1, and osteoblast as alveolar bone regeneration in socket preservation. Department of Prosthodontic, Faculty of Dental Medicine, Universitas Airlangga, Surabaya © 2018 The Journal of Indian Prosthodontic Society | Published by Wolters Kluwer Medknow 60132, Indonesia.
- Stokes, Debbie J. (2008). Principles and Practice of Variable Pressure Environmental Scanning Electron Microscopy (VP-ESEM). Chichester: John Wiley & Sons. ISBN 978-0470758748.
- Shi B., Zhou Y., Wang YN., Cheng XR. Alveolar ridge preservation prior to implant placement with surgicalgrade calcium sulphate and platelet-rich plasma: A pilot study in a canine model. Int J Oral Maxillofac Implants 2007; 22:656-665.
- Margarida Figueiredo, Jose Henriques, Gabriela Martins, Fernando Guerra, Fernando Judas, Helena Figueiredo. Physicochemical Characterization of Biomaterials Commonly Used in Dentistry as Bone Substitutes— Comparison with Human Bone. Received 27 October 2008; revised 30 May 2009; accepted 27 July 2009. Published online 10 November 2009 in Wiley InterScience (www. interscience.wiley.com). DOI: 10.1002/jbm.b.31529
- Ichiro H., DDS, Eriko M., DDS, PhD, Yukinobu T., DDS, PhD, and Ken O., DDS, PhD. Effects of Platelet-Poor Plasma, Platelet-Rich Plasma, and Platelet-Rich Fibrin on Healing of Extraction Sockets with Buccal Dehiscence in Dogs TISSUE ENGINEERING: Part A Volume 20, Numbers 3 and 4, 2014.
- Rothamel D., Schwarz F., Herten M., Engelhardt E., Donath K., Kuehn P., Becker J.: Dimensional ridge alterations following socket preservation using a nanocrystalline hydroxyapatite paste. A histomorphometrical study in dogs. Int. J. Oral Maxillofac. Surg. 2008; 37: 741–747
- Lekovic V., Camargo PM., Klokkevold PR., et al. Preservation of alveolar bone in extraction sockets using bioabsorbable membranes. J Periodontol 1998;69: 1044-1049.

- Vance GS., Greenwell H., Miller RL., Hill M., Johnston H., Scheetz JP. Comparison of an allograft in an experimental putty carrier and a bovine-derived xenograft used in ridge preservation: A clinical and histologic study in humans. Int J Oral Maxillofac Implants 2004;19:491-497.
- Chaitanya P., Cynthia B., Nitin H. Comparative Alveolar Ridge Preservation Using Allogenous Tooth Graft versus Free-dried Bone Allograft: A Randomized, Controlled, Prospective, Clinical Pilot Study. Contemp Clin Dent. 2017; Aprl- Jun; 8(2): 211-217.
- Nevins, M., Camelo, M., De Paoli, S., Friedland, B., Schenk, R. K., Parma-Benfenati, S.,Simion, M., Tinti, C. & Wagenberg, B. A study of the fate of the buccal wall of extraction sockets of teeth with prominent roots. International Journal of Periodontics and Restorative Dentistry 26, 19–29, 2006.
- Araujo M, Linder E, Wennstro m, J. & Lindhe, J. The influence of Bio-Oss collagen on the healing of an extraction socket:an experimental study in the dog. International Journal of Periodontics and Restorative Dentistry 28, 123–135, 2008.
- Wang HL & Tsao, Y. Histologic evaluation of socket augmentation with mineralized human allograft. International Journal of Periodontics and Restorative Dentistry 28, 231–237, 2008.
- Kassem M, Blum W, Ristelli J, Mosekilde L, Eriksen EF. Growth hormone stimulates proliferation and differentiation of normal human osteoblast-like cells in vitro. Calcif Tissue Int 1993; 52: 222-6.
- Hogan BL. Bone morphogenic proteins: Multifunctional regulators of vertebrate development. Genes Dev 1996; 10: 1580-94.
- Hayden JM, Mohan S, Baylink DJ. The insulin-like growth factor system and the coupling of formation to resorption. Bone 1995; 17(Suppl 2): 93S-8S.
- Goldstein J, Newburry D, Joy D, Lyman C, Echlin P, Lifshin E et al. Scanning electron microscopy and x-ray microanalysis. 3rd New York: Plenum Press; 2003.
- Joseph G., Newbury E., Joy D., Lyman C. Scanning Electron Microscopy and X-ray Microanalysis. Springer, 3 (2003). Available at: http://works.bepress.com/joseph_ goldstein/15/
- Fickl S, Zuhr O, Wachtel H, Bolz W & Huerzeler MB, (2008) Hard tissue alterations after socket preservation: an experimental study in the beagle dog. Clinical Oral Implants Research 19: 1111–1118.

- 22. Zhang W, Walboomers X, van Osch G, van den Dolder J, Jansen J. (2008) Hard tissue formation in a porous HA/ TCP ceramic scaffold loaded with stromal cells derived from dental pulp and bone marrow. Tissue Eng Part A 14:285.
- Farina NM, Guzon FM, Pena ML, Cantalapiedra AG. (2008) In vivo behaviour of two different biphasic ceramic implanted in mandibular bone of dogs. J Mater Sci Mater Med 19:1565–1573.
- Cordonnier T., Layrolle P., Gaillard J., Langonne A., Sensebe L., Rosset P et al. (2010) 3D environment on human mesenchymal stem cells differentiation for bone tissue engineering. J Mater Sci Mater Med 21:981–987.
- Cordonnier T., Langonne A., Corre P., Renaud A., Sensebe L., Rosset P et al. (2014) Osteoblastic differentiation and potent osteogenicity of three-dimensional hBMSC-BCP particle constructs. J Tissue Eng Regen Med 8:364–376.
- Yang C., Unursaikhan O., Lee JS., Jung UW., Kim CS., Choi SH. (2014) Osteoconductivity and biodegradation of synthetic bone substitutes with different tricalcium phosphate contents in rabbits. J Biomed Mater Res B Appl Biomater 102:80–88.
- Pripatnanont P., Praserttham P., Suttapreyasri S., Leepong N., Monmaturapoj N. (2016) Bone regeneration potential of biphasic nanocalcium phosphate with high hydroxyapatite/ tricalcium phosphateratios in rabbit calvarial defects. Int J Oral Maxillofac Implant 31:294–303.
- 28. Ebrahimi M., Botelho M., Dorozhkin S. Biphasic calcium phosphates bioceramics (HA/TCP): concept, physicochemical properties and the impact of standardization of study protocol sinbiomaterials research, Mater.Sci.Eng.C(2016).
- Ebrahimi M., Pripatnanont P., Suttapreyasri S., Monmaturapoj N.. Fabrication and characterization of novel nano hydroxyapatite/β-tricalcium phosphate scaffolds in three different composition ratios. J Biomed Mater Res A.100 (9) (2012), 2260-2268.
- 30. Dorozhkin S. Biphasic,triphasic and multiphasic calcium orthophosphates. Acta Biomater.8(2012) 963–977.
- Fountos G., Yasumura S., Glaros D.: The skeletal calcium/ phosphorus ratio: a new in vivo method of determination. Med. Phys. 24, 1303-1310 (1997). doi:10.1118/1.598152
- 32. Fountos, G., Kounadi, E., Tzaphlidou, M., Yasumura, S., Glaros, D.: The effects of inflammationmediated

- osteoporosis (IMO) on the skeletal Ca/P ratio and on the structure of rabbit bone and skin collagen. Appl. Radiat. Isotopes 49, 657-659 (1998).
- Fountos G., Tzaphlidou M., Kounadi E., Glaros D.: In vivo measurement of radius calcium/phosphorus ratio by X-ray absorptiometry. Appl. Radiat. Isotopes 51, 273-278 (1999). doi:10.1016/S0969-8043(99)00056-1
- Margaret T.: Bone Architecture: Collagen Structure and Calcium/Phosphorus Maps. J Biol Phys (2008) 34:39-49 DOI 10.1007/s10867-008-9115-y
- Brodie J., Merry J., Grant MH. The mechanical properties of calcium phosphate ceramics modified by collagen coating and populated by osteoblasts. J Mater Sci Mater Med 2006;17:43–48.
- 36. Yasuhiro T., Yo S., Asuka M., Takashi M., Norihiro N.: Effect of Varying HAP/TCP Ratios in Tape-cast Biphasic Calcium Phosphate Ceramics on Responce In Vitro. Journal of Hard Tissue Biology (2009) 71-76.
- 37. Mehdi E., Prisana P., Naruporn M., Srisurang S.: Fabrication and characterization of novel nano hydroxyapatite/b-tricalcium phosphate scaffolds in three different composition ratios. Journal of Biomedical Materials Research (2012). VOL 100A (9).
- 38. Ying C., JingW., Xiangdong Z., Xuening C., Xiao Y., Kai Z., Yujiang F., Xingdong Z.: The directional migration and differentiation of mesenchymal stem cells toward vascular endothelial cells stimulated by biphasic calcium phosphate ceramic. Regenerative Biomaterials, 2018, 129–139.
- 39. Ducheyne P.; Qiu, Q. Bioactive ceramics: The effect of surface reactivity on bone formation and bone cell function. Biomaterials 1999, 20, 2287–2303.
- 40. Rompen E., Domken O., Degidi M., Farias P., A.E., Piattelli, A. The effect of material characteristics, of surface topography and of implant components and connections on soft tissue integration: A literature review. Clin. Oral Implants Res. 2006, 17, 55–67.
- Bajpai, P., Benghuzzi H.. Ceramic systems for long delivery of chemicals and biological. J. Biomed. Mater. Res. 1988, 22, 1245–1266.
- 42. Ren W., Song W., Esquivel A., Jackson N., Nelson M., Flynn J., Markel D. Effect of erythromycin-doped calcium polyphosphate scaffold composite in a mouse pouch infection model. J. Biomed. Mater. Res. Part B Appl. Biomater. 2014, 102, 1140–1147.

(188)

- Aneja A., Woodall J., Wingerter S., Tucci M., Benghuzzi H. Analysis of tobramycin release from beta tricalcium phosphate drug delivery system. Biomed. Sci. Instrum. 2008, 44, 88–93.
- Gao S., Shiota M., Fujii M., Chen K., Shimogishi M., Sato M., Kasugai S. Combination of simvastatin and hydroxyapatite fiber induces bone augmentation. Open J. Regen. Med. 2013, 2, 53–60.
- 45. Muñoz F., López-Peña M., Miño N., Gómez-Moreno G., Guardia J., Cutando A. Topical application of melatonin and growth hormone accelerates bone healing around dental implants in dogs. Clin Implant Dent Relat Res. 2012 Apr; 14 (2):226-35.
- Calvo-Guirado J., Mate-Sanchez J., Delgado-Ruiz R., Ramirez-Fernández M., Cutando-Soriano A., Peña M. Effects of growth hormone on initial bone formation around dental implants: a dog study. Clin Oral Implants Res. 2011 Jun; 22(6):587-93.
- 47. Hossam E., Elghamrawy S., Osman S., Elhak A. Histological evaluation of the effect of using growth hormone around immediate dental implants in fresh

- extraction sockets: an experimental study. Implant Dent. 2011 Feb; 20(1):47-55.
- 48. Ko-Ning H., Eisner S., Kuo-Chi C., Tsai-Chin S., Yu-Chi C., Haw-Ming H., Nai-Chia T., Che-T., Sheng-W., Wei-Jen C. A Novel HA/β-TCP-Collagen Composite Enhanced New Bone Formation for Dental Extraction Socket Preservation in Beagle Dogs. Materials 2016, 9, 191; 1-15.
- 49. Rander M., MSc, Suzie A., PhD, José A., PhD, Luiz G., PhD. Bone Integration Behavior of Hydroxyapatite/b-Tricalcium Phosphate Graft Implanted in Dental Alveoli: A Histomorphometric and Scanning Electron Microscopy Study. Implant Dentistry 2014; 23 (6): 710-715.
- Kitsugi T., Yamamuro T., Nakamura T., et al. Transmission electron microscopy observations at the interface of bone and four types of calcium phosphate ceramics with different calcium/phosphorus molar ratios. Biomaterials. 1995;16:1101–1107.
- Fujita R., Yokoyama A., Nodasaka ., et al. Ultrastructure of ceramic-bone interface using hydroxtapatite and beta-tricalcium phosphate ceramics and replacement mechanism of beta-tricalcium phosphate in bone. Tissue Cell. 2003;35:427–440.