



The Effect of Titanium Dioxide Nanoparticles Against Cyanobacterial Microorganisms

Karima Boutarfa^{1,2*}, Amal Saoudi¹, Khedidja Boufligha¹, Houneida Benbouzid³,
Racha Mihoub², Mourad Bensouilah¹

¹- Ecobiology Laboratory for Marine Environments and Costal Areas, Marine Sciences Departement , Badji Mokhtar-Annaba University, 23000 ALGERIA.

²- Biochimistry and Applied Microbiology Laboratory, Biochimistry Department, Faculty of sciences, Badji Mokhtar- Annaba University, 23000 ALGERIA.

³- Cellular Toxicology Laboratory, Biology Departement, Faculty of sciences, Badji Mokhtar- Annaba University, 23000 ALGERIA.

*Corresponding Author: karima.biob@yahoo.fr

ARTICLE INFO

Article History:

Received: March 18, 2021

Accepted: April 18, 2021

Online: May 25, 2021

Keywords:

Antibacterial effect,
Titanium dioxide,
nanoparticles (TiO₂ NPs),
Cyanobacteria,
Microcystis sp.,
Health risks.

ABSTRACT

Titanium dioxide nanoparticles (TiO₂ NPs) present a high interest as an excellent antibacterial agent against pathogenic microorganisms. Currently, the massive proliferation of potentially toxic cyanobacteria in the water of dams presents a threat to the public health of these water consumers. This study was maintained to evaluate the antibacterial activity of TiO₂ NPs against the cyanobacteria « *Microcystis* sp.» responsible for toxic blooms in the freshwater reservoirs intended for drinking water production. Three concentrations of TiO₂ NPs (150 mg.l⁻¹, 300 mg.l⁻¹ and 600 mg.l⁻¹) were tested in the laboratory on two strains of *Microcystis* sp. (S1, S2) over a period of eight days. In parallel, three biotic variables were measured: *Microcystis* sp. cells density (cell.ml⁻¹), Chlorophyll (a) (µg.l⁻¹), and Phycocyanin (µg.l⁻¹) content every 48 hours all along the experimental period. A remarkable decrease in cell densities was registered, recording decreased values from 77490 to 39091 cell.ml⁻¹ for S1 and from 2801 to 0 cell.ml⁻¹ for S2 with the concentration of 600 mg.l⁻¹. Additionally, the chlorophyll (a) content fell from 45 to 8 µg.l⁻¹ for S1, and from 34 to 0 µg.l⁻¹ for S2. Similarly, treatment with TiO₂ NPs caused the degradation of phycocyanin from 200 to 65 µg.l⁻¹ for S1 and from 200 to 0 µg.l⁻¹ for S2 with the highest concentration of these nanoparticles (600 mg.l⁻¹). TiO₂ NPs exhibit a very important antibacterial effect against *Microcystis* sp. Thus, TiO₂ NPs can be used in the future to reduce the high biomasses of cyanobacteria during the process of tap water production.

INTRODUCTION

The innovative nanotechnology applications and their nanomaterials are primarily used in biomedical, environmental and electronic fields. The group of metal oxides nanoparticles is the largest group of nanomaterials that received a significant attention being, hence, applied in a plenty of products.

Obeizi *et al.* (2020) and **Obeizi *et al.* (2021)** confirmed that, several metal oxides might show bacteriostatic or bactericidal effect. This antibacterial activity differs according to the type and number of the tested bacteria, depending on the physicochemical properties, the concentration and the experimental conditions of the used nano-molecules (**Vojislav *et al.*, 2020**).

Titanium dioxide nanoparticles (TiO₂ NPs) are among the most globally produced nanomaterials (**Piccino *et al.*, 2012**) due to their high refractive index and resistance to discolouration, especially to stability, photocatalytic, depolluting and antibacterial properties. **Kahru and Dubourguier (2010)** classified those NPs as harmful to many water organisms such as crustaceans and nematodes. Moreover, they may harm microorganisms as yeast, micro-algae and bacteria. Considerably, NPs are used as catalyst for water purification (**Botelho *et al.*, 2014; Janer *et al.*, 2014**) and as an antimicrobial agent as well (**Benbouzid *et al.*, 2019**).

On the other hand, the increasing cyanobacterial bloom occurrences around the world has introduced a considerable research subject for scientists (**Merel *et al.*, 2013**) to identify the responsible factors for their proliferation. Additionally, this may draw scientist to think carefully and place in action an arranged and well structured surveillance system that can prevent and reduce the cyanobacterial bloom occurrence. Such a concern is due to the toxicological risks of those microorganisms and the impact of which that uncontrolled cyanobacterial blooms may have on aquatic ecosystems (**Carman & Tomevska, 2019**).

It is worth mentioning that, the cyanobacteria degrade water quality (**Fleming *et al.*, 2002**) and produce several cyanotoxins as neurotoxins, hepatotoxins, and dermatotoxins that can affect both animals and humans (**Lance *et al.*, 2010a, b; Carmichael & Boyer, 2016 ; Metcalf *et al.*, 2020**). In this context, the frequent consumption and/or the exposure to the contaminated drinking water or aquatic foods can lead to a serious threat to public health safety (**Agasild *et al.*, 2019**).

Consequently, the main aim of this study was to evaluate the antibacterial effect of TiO₂ NPs on a potentially toxic cyanobacteria on basis of the probability of using those NPs in water treatment processes in the future. The present test was achieved via the exposure of two freshwater strains *Microcystis* sp. on an increasing concentrations of TiO₂ NPs suspension, where three biotic parameters were measured (*Microcystis* sp. cells density (cell.ml⁻¹), Chlorophyll (a) (µg.l⁻¹) and Phycocyanin (µg.l⁻¹) contents) in a period of eight days.

MATERIALS AND METHODS

The preparation of TiO₂ NPs suspension

Three concentrations of TiO₂ NPs (150 mg.l⁻¹, 300 mg.l⁻¹ and 600 mg.l⁻¹) were prepared in BG11 medium (particules were kindly donated by the Surfaces and Solids

Interfaces Laboratory Badji mokhtar-Annaba University), then the solutions were sonicated with pulsatile movements for 5min to insure the dispersion of the nanomolecules.

Cyanobacteria culture and cells growth testing

The growth of cyanobacterial cells culture was maintained in BG11 medium, pH = 7 at 25 °C with a photoperiodic cycle of 12h: 12h light and dark. Each culture contained 1g of *Microcystis* sp. cells (*Microcystis* sp. strains were collected from Algerian freshwaters in 2019) then the prepared concentrations were added (150 mg.l⁻¹, 300 mg.l⁻¹ and 600 mg.l⁻¹ of TiO₂ NPs) to the *Microcystis* sp. Cultures. A frequent agitation was realized to avoid precipitation of any cell or molecule in the flasks. Each prepared serie contained a control sample without NPs. The experience extended for eight days successively.

Cells densities was counted with "Nageotte" counting cell using a Carl Zeiss model (Axiostar plus) microscope equipped with digital uEye32 camera and according to the technique of **Brient *et al.* (2001)**. The aforementioned work was achieved beginning from day 0, every 48 hours all along the experiment. To minimize error, the counting operation was performed in triplicate. The cell densities were expressed in (cells.ml⁻¹)

Photosynthetic pigments measurement

The chlorophyll (a) (Chl a) pigments were measured using chlorophyll TRIOS MicroFlu-Chl probe; which uses a fluorimeter equipped with a blue diode. The excitation wave length filter is at 470 nm and the emission is at 685 nm, with a range from 0 to 100 µg.l⁻¹ and a sensitivity = 0.1 µg.l⁻¹. The results were expressed in µg.l⁻¹.

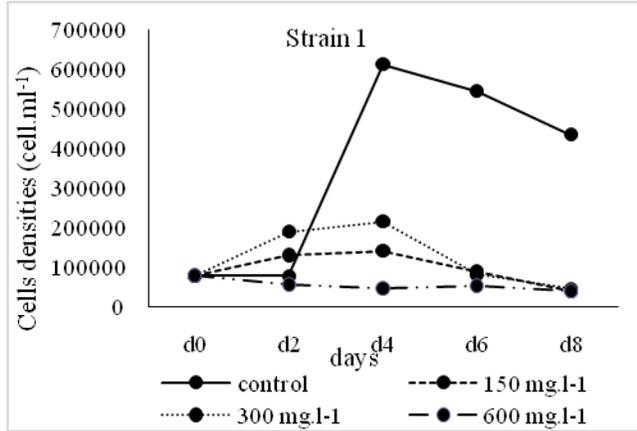
Phycocyanin (PC) pigments (pigment characteristic of cyanobacteria) measurement was carried out using a sensitive and high selective probe (TRIOS MicroFlu-blue phycocyanin) which uses a fluorimeter, submersible and miniaturized (diameter of 48 mm and 200 mm length), equipped with an ultra-brightred LEDs, and provided with excitation wave length filter at 620 nm and emission at 655 nm with a band width of 10 nm. This probe gives a linear response to phycocyanin concentration up to 200 µg.l⁻¹ and a precision of 0.02 µg.l⁻¹. The results were expressed in µg.l⁻¹.

RESULTS

***Microcystis* sp. densities variation**

The contact between *Microcystis* sp. strains and the TiO₂ NPs allowed a reduction in the densities which varies from 213926 cells.ml⁻¹ to 38441 cells.ml⁻¹ for S1 and from 15959 cells.ml⁻¹ to 0 cells.ml⁻¹ for S2. Fig.(1) shows a decreasing variation in the function of TiO₂ NPs concentrations and the exposure time to those nanoparticules, with a remarkable reduction in S2 from the second day comparing to S1. The highest TiO₂ NPs concentration gave the highest rate of cell decrease of S1 after 8 days of contact, however, the S2 was eliminated by the 150 mg.l⁻¹ dose from the second day. Those results

show that the TiO₂ NPs exerts a good bactericidal activity against the two strains of *Microcystis* sp., and that the antibacterial activity is directly linked to the concentration of TiO₂ and the contact time. Furthermore, it was noticed that the S2 was more sensitive than the S1.



(cells.ml⁻¹) with the three TiO₂ NPs concentrations during the experimental period.

Fig. 1. *Microcystis* sp. cells density

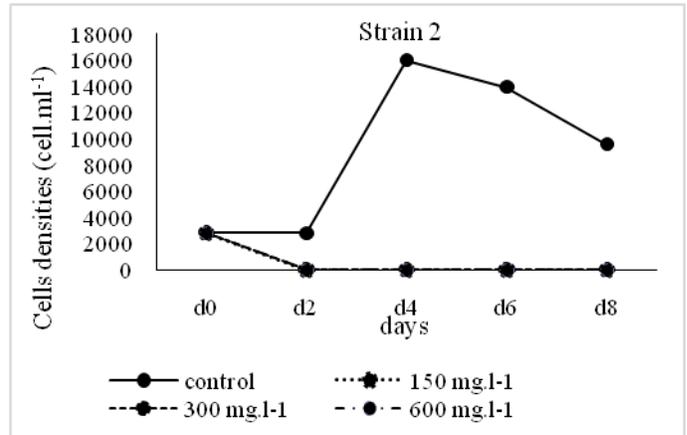


Fig.(2) shows a representative explanation for the damages that TiO₂ NPs caused to the cells of *Microcystis* sp. in particular, and to the colonies in general. The mucilage membrane is deformed and the molecules of TiO₂ NPs are clearly covering the colonies.

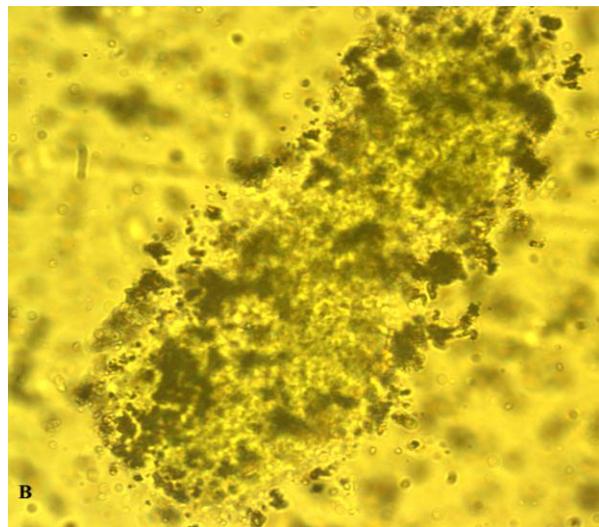


Fig. 2. *Microcystis* sp. colonies apparition under (Axiostar plus) microscope GRx400, A : Control solution, B : Presence of TiO₂ NPs (2019).

Photosynthetic pigments variation

The results of the chlorophyll (Chl) (a) and phycocyanin content in the two strains; S1 and S2, represented in Fig. (3 & 4) show a decrease that is proportional to the three concentrations of TiO₂ NPs and the contact time. The lowest content of the two pigments were observed in S1, with the concentration of 600 mg.l⁻¹ after eight days of contact. However, the lowest values for S2 were observed with the concentration of 150 mg.l⁻¹ beginning from the second day. The maximum concentration of Chl(a) content was registered with the S1 (45 µg.l⁻¹), however the minimum concentration was 0 µg.l⁻¹ for the second one. Thus, the levels of PC varied from 200 µg.l⁻¹ to 65 µg.l⁻¹ for S1 while for the second strain, they ranged from 200 µg.l⁻¹ to 0 µg.l⁻¹.

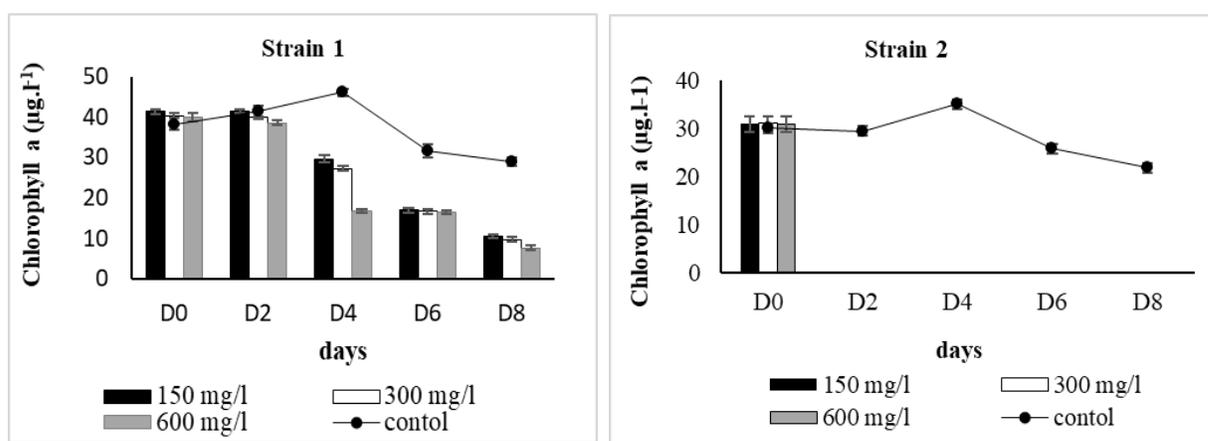


Fig. 3. Chlorophyll (a) concentrations (µg.l⁻¹) on the function of the TiO₂ NPs concentrations along the eight days.

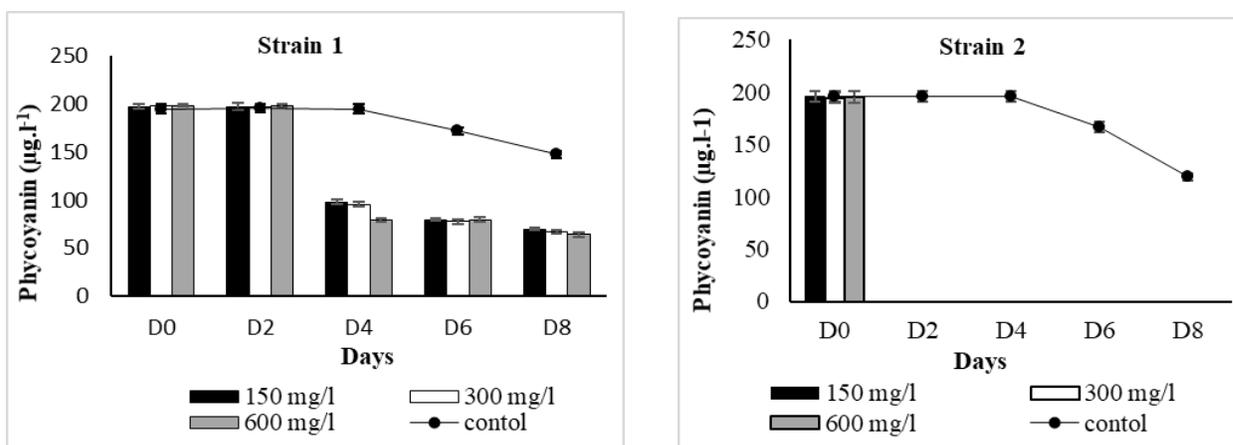


Fig. 4. Phycocyanin pigments concentration (µg.l⁻¹) on the function of the TiO₂ NPs concentrations along the eight days.

DISCUSSION

The results relating to both of the cyanobacterial strains densities showed a proportional decrease with the tested concentrations of TiO₂ NPs. The lowest densities were observed with the highest concentration (600 mg.l⁻¹). Microscopic examination allowed researchers to see the antibacterial effect of TiO₂ NPs on *Microcystis* sp. through the deformation in the morphology of cell walls and colonies, besides the presence of a large number of destroyed cells in the slide (Fig. 2).

The membrane of cyanobacteria is charged negatively, it is composed of peptidoglycan, glycopeptide and polysaccharide which offers many binding sites for TiO₂ NPs since it is charged positively via non-specific interactions ; hydrogen bonding, hydrolytic or electrostatic (Rai *et al.*, 2009; Liu *et al.*, 2018). Those bonds can perforate cells membrane and allow the accumulation of those nanoparticles inside the cell. The nanoparticles accumulation would, hence, generate oxidative stress (Tran *et al.*, 2016; Hu *et al.*, 2018). Consequently, the intensive presence of radical molecules (OH[•], O₂⁻) would cause non reversible damage that leads to cell death.

Rai *et al.* (2009) reported that the smallest nanoparticles have the strongest bactericidal effect. Thus, the antibacterial effect of TiO₂ as a metal oxide nanoparticles on *Microcystis* sp. strains is probably due to the small size that allows their penetration inside the cell causing oxidative stress via over production of reactive oxygen species « ROS ».

In addition, Miller *et al.* (2012) reported that, the « ROS » high production is due to protein oxidation, unsaturated lipids or even nucleicacids, which can cause serious DNA damage and may lead to bacterial cell destruction. Noticeably, TiO₂ dissolution to Ti⁺² and OH[•], in the presence of water and oxygen, can lead to the formation of a highly reactive hydroxyl radical which can react with 2-deoxyribose of DNA and generate a modified nucleic bases. The generated element is one of the predominant lesions generated by the hydroxyl radical in the body that corresponds to the 8-oxo-7,8-dihydroguanine resulting from the oxidation of guanine by TiO₂ NPs.

For the two photosynthetic pigments: the results of both Chlorophyll (a) and phycocyanin showed a decrease with the nano-concentrations, in a way that coincides with results of Liu *et al.* (2018) and Tran *et al.* (2016). Chl (a) reduction can be explained by the presence of the O₂⁻ radical which destroys that pigment. Based on those parameters, it was found that, TiO₂ NPs exerted an antibacterial effect against *Microcystis* sp. with respect to the two strains S1 and S2.

CONCLUSION

It can be concluded from the results of the present study that TiO₂ NPs have an antibacterial effect on *Microcystis* sp. and that this effect is directly dependent on the concentrations of TiO₂ NPs, the contact time with *Microcystis* sp. and susceptibility or resistance of the strains. Therefore, TiO₂ NPs can be offered as an antibacterial agent against *Microcystis* sp. with doses that do not exceed 600 mg.l⁻¹.

With a view to reduce health and economic risks associated with the presence of *Microcystis* sp. and its toxins (Microcystin LR) in the majority of blooms produced in drinking water production, further investment is recommended to know at which stage of water treatment it should be introduced. Nevertheless, the current study should be supplemented by eco-toxicological studies on TiO₂ NPs to prevent their approximate risks on environmental and public health.

ACKNOWLEDGEMENTS

This research did not receive any specific grant from funding agencies in the public, commercial, or not-for-profit sectors.

REFERENCES

- Agasild, H.; Panksep, K.; Tonno, I.; Blank, K.; Koiv, T.; Freiberg, R.; Laugaste, R.; Jones, R.I.; Noges, P. and Noges, T.** (2019). Role of potentially toxic cyanobacteria in crustacean zooplankton diet in a eutrophic lake. *Harmful Algae.*, 89: 101688.
- Benbouzid, H.; Obeizi, Z. and Djahoudi, A.E.G.** (2019). Antimicrobial Effects of TiO₂ Nanoparticles against Drug-Resistant Bacteria and *Candida albicans*. *IJIAAR.*, 3(2) : 177-182.
- Botelho, M.C.; Costa, C.; Silva, S.; Costa, S.; Dhawan, A.; Oliveira, P.A. and Teixeira, J.P.** (2014). Effects of titanium dioxide nanoparticles in human gastric epithelial cells in vitro. *Biomed. Pharmacother.*, 68(1): 59-64.
- Brient, L.; Vézic, C. and Bertru, G.** (2001). Evaluation des efflorescences à cyanobactéries dans des eaux de cours d'eau et plans d'eau breton. Université de Rennes 1., 80pp.
- Carman, R. and Tomevska, S.** (2019). A Million Fish Dead in 'distressing' Outback Algal Bloom at Menindee.
- Carmichael, W.W.; and Boyer, G.L.** (2016). Health impacts from cyanobacteria harmful algae blooms: Implications for the North American Great Lakes. *Harmful*

Algae., 54: 194–212.

- Fleming, L.E.; Rivero, C.; Burns, J.; Williams, C.; Bean, J.A.; Shea, K.A. and Stinn, J.** (2002). Blue green algal (cyanobacterial) toxins, surface drinking water, and liver cancer in Florida. *Harmful Algae.*, 1(2): 157-168.
- Hu, J.; Wang, J.; Liu, S.; Zhang, Z.; Zhang, H; Cai, X.; Pan, J. and Liu, J.** (2018). Effect of TiO₂ nanoparticule aggregation on marine microalgae *isochrysis galbana*. *J. Environ.Sci.*, 66: 208– 215.
- Janer, G.; del Molino, E.M.; Fernández-Rosas, E.; Fernández, A. and Vázquez-Campos, S.** (2014). Cell uptake and oral absorption of titanium dioxide nanoparticles. *Toxicol Lett.*, 228(2): 103-110.
- Kahru, A. and Dubourguier, H.C.** (2010). From ecotoxicology to nanoecotoxicology. *Toxicology.*, 269: 105-119.
- Lance, E.; Josso, C.; Dietrich, D.; Ernst, B.; Paty, C.; Senger, F.; Bormans, M.; and Gerard, C.** (2010)a. Histopathology and microcystin distribution in *Lymnaea stagnalis* (*Gastropoda*) following toxic cyanobacterial or dissolved microcystin-LR exposure. *Aquat. Toxicol.*, 98: 211–220.
- Lance, E.; Brient, L.; Carpentier, A.; Acou, A.; Marion, L.; Bormans, M.; and Gérard, C.** (2010)b. Impact of toxic cyanobacteria on gastropods and microcystin accumulation in a eutrophic lake (Grand-Lieu, France) with special reference to *Physa* (= *Physella*) *acuta*. *Sci. Total Environ.*, 408: 3560–3568.
- Liu, J; Pinghe, Y. and Zhao, L.** (2018). Adverse effect of nano-TiO₂ on the marine macroalgae *Gracilaria lemaneiformis* (*Gracilariales*, *Rhodophyta*): growth and antioxidant activity. *China RSC Adv.*, 8: 29172–29178.
- Merel, S.; Walker, D.; Chicana, R.; Snyder, S.; Baures, S. and Thomas, O.** (2013). State of knowledge and concerns on cyanobacteria blooms and cyanotoxins. *Environ. Int.*, 59: 303–327.
- Metcalf, J.S. and Codd, G.A.** (2020). Co-Occurrence of Cyanobacteria and Cyanotoxins with Other Environmental Health Hazards : Impacts and Implications. *Toxins.*, 12: 629.
- Miller, R.J.; Bennett, S.; Keller, A.A.; Pease, S. and Lenihan, H.S.** (2012). TiO₂ nanoparticles are phototoxic to marine phytoplankton. *PLoS One.*, 7(1): 30321.
- Obeizi, Z.; Benbouzid, H.; Ouchenane, S.; Yilmaz, D.; Culhad, M. and Bououdina, M.** (2020). Biosynthesis of Zinc oxide nanoparticles from essential oil of *Eucalyptus globulus* with antimicrobial and anti-biofilm activities. *Mater. Today Commun.*, 25: 1-10.

-
- Obeizi, Z.; Benbouzid, H.; Bouarroudj, T.; Benzaid, C. and Djahoudi, A.** (2021). Synthesis, characterization of (Ag-SnO₂) nanoparticles and investigation of its antibacterial and anti-biofilm activities. *J. New Technol. Mater.*, 10(2): 10-17
- Piccinno, F.; Gottschalk, F.; Seeger, S. and Nowack, B.** (2012). Industrial production quantities and uses of ten engineered nanomaterials in Europe and the world. *J Nanopart Res.*, 14(9): 1-11.
- Rai, M.; Yadav, A. and Gade, A.** (2009). Silver nanoparticles as a new generation of antimicrobials. *Biotechnol. Adv.*, 27: 76-83.
- Tran, T.H.; Nguyen, T.K.; Nguyen, T.T.; Phuong, T.; Le, Thi P.; Do, V.; Trinh, Q.H. and Duong, T.T.** (2016). Nanoparticles as a control for cyanobacterial bloom. *J. Viet. Env.*, 8(3): 161-166.
- Vojislav, S.; Sladjana, B. and Tanasković.** (2020). Antibacterial activity of metal oxide nanoparticles. In: "Micro and Nano Technologies". Rajendran, S.; Mukherjee, A.; Nguyen, T.A.; Godugu, C.; Shukla, R.K. (Eds.). Nanotoxicity., pp.241-274.