Egyptian Journal of Aquatic Biology & Fisheries Zoology Department, Faculty of Science, Ain Shams University, Cairo, Egypt. ISSN 1110 – 6131 Vol. 23(1): 299 -304 (2019)



# Bacterial diversity and distribution in Soft Corals and Sponges in the Red Sea

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#### **ARTICLE INFO**

www.ejabf.journals.ekb.eg

# **Article History:**

Received: Jan.23, 2019 Accepted: Feb. 26, 2019 Online: March 2019

#### **Keywords**:

Bacteria Diversity Soft Corals Sponges Red Sea

#### **ABSTRACT**

The Red Sea has high biodiversity, with at least 266 coral and sponge species. The coral/Sponge holobiont is comprised of the coral/sponge animal and its associated microorganisms consisting of bacteria, archaea, fungi and viruses. It has been hypothesized that this interaction plays a role in coral/ sponge defense. This study aims to study the bio-diversity of some collected types of sponge and soft coral from Red Sea and to isolate associated bacteria from them. The result includes five types of sponges; *Ircinia strobilina*, *Callyspongia viridis*, *Suberea* spp., *Spongia officinalis* and *Biemna ehrenbergi* and five types of soft corals; *Heteroxenia fuscescens*, *Didemnum moseleyi*, *Lobophytum pauciflorum*, *Sarcophyton trocheliophorum* and *Sinularia* spp. Twenty two bacterial isolates were isolated from the sponges with 41 % and thirty two bacterial isolates were isolated from soft coral samples with 59 % collected from different Red Sea areas.

#### INTRODUCTION

Coral reefs are well developed along Red Sea which has a coast extend to 2270 Km (Kotb et al., 2008). It contains biologically diverse sponges and corals and considered to be among the most diverse in the world species (Loya, 1972). These corals and sponges show contrasts in their biology and environmental attributes (Sheppard et al., 1992). They are considered economically valuable, providing shelter, food and breeding sites for plants and animals living in oceans (Rinkevich, 2005). Sponges and coral reefs are considered to be natural ecosystem and provide an excellent habitat for marine organisms due to their structure and high retention of nutrients (Stoeckl et al., 2014). Bacteria and other microorganisms are ubiquitous in the marine environment are taxonomically diverse and biologically active (Rheinheimer, 1992). Marine organisms such as corals and sponges are mostly colonized by bacteria and the surface of living corals is covered by mucus which is colonized by bacteria, allowing for the establishment of diverse bacterial community, some of these bacteria can be pathogenic and some of them can be beneficial (Ritchie et al., 2017). So this study aims to study the bio-diversity of some sponge and soft coral collected from Red Sea.







#### MATERIALS AND METHODS

# Soft coral/Sponge collection and cultivation condition

From three different areas in Red Sea, Egypt, in depth of  $\pm$  5-8m, sponge and soft corals sampled were collected and transferred into a sterilized plastic bag under the water itself and stored in the ice box and transferred to the laboratory for identification and isolation of the associated bacteria.

## Isolation of sponge and soft coral associated bacteria (Chen et al., 2012)

Sponge and soft corals samples were washed with autoclaved filtered sea water. Sponges and soft corals were cut into small pieces and were homogenized by grinding using sterile mortar and sterile sea water.

Table 1: biochemical test used for bacterial identification using VITEK 2

2 1: biochemical test used for bacterial identification using VITEK 2.  Gram Positive tests  Gram Positive testes		
Ala-Phe-Pro-ARYLAMIDASE	D-AMYGDALIN	
ADONITOL	PHOSPHATIDYLINOSITOL PHOSPHOLIPASE (	
	D-XYLOSE	
L-Pyrrolydonyl-ARYLAMIDASE		
L-ARABITOL	ARGININE DIHYDROLASE 1	
D-CELLOBIOSE	BETA-GALACTOSIDASE	
BETA-GALACTOSIDASE	ALPHA-GLUCOSIDASE	
H2S PRODUCTION	Ala-Phe-Pro ARYLAMIDASE	
BETA-N-ACETYL-GLUCOSAMINIDASE	CYCLODEXTRIN	
Glutamyl Arylamidase pNA	L-Aspartate ARYLAMIDASE	
D-GLUCOSE	BETA GALACTOPYRANOSIDASE	
GAMMA-GLUTAMYL-TRANSFERASE	ALPHA-MANNOSIDASE	
FERMENTATION/ GLUCOSE	PHOSPHATASE	
BETA-GLUCOSIDASE	Leucine ARYLAMIDASE	
D-MALTOSE	L-Proline ARYLAMIDASE	
D-MANNITOL	BETA GLUCURONIDASE	
D-MANNOSE	ALPHA-GALACTOSIDASE	
BETA-XYLOSIDASE	L-Pyrrolydonyl-ARYLAMIDASE	
BETA-Alanine arylamidase	BETA-GLUCURONIDASE	
L-Proline ARYLAMIDASE	Alanine ARYLAMIDASE	
LIPASE	Tyrosine ARYLAMIDASE	
PALATINOSE	D-SORBITOL	
	UREASE	
Tyrosine ARYLAMIDASE		
UREASE URE	POLYMIXIN B RESISTANCE	
D-SORBITOL	D-GALACTOSE	
SACCHAROSE/SUCROSE	D-RIBOSE	
D-TAGATOSE	L-LACTATE alkalinization	
D-TREHALOSE	LACTOSE	
CITRATE (SODIUM)	N-ACETYL-D-GLUCOSAMINE	
MALONATE	D-MALTOSE	
5-KETO-D-GLUCONATE	BACITRACIN RESISTANCE	
L-LACTATE alkalinization	NOVOBIOCIN RESISTANCE NOVO	
ALPHA-GLUCOSIDASE	GROWTH IN 6.5% NaCl	
SUCCINATE alkalinization	D-MANNITOL	
Beta-N-ACETYL-GALACTOSAMINIDASE	D-MANNOSE	
ALPHA-GALACTOSIDASE	METHYL-B-D-GLUCOPYRANOSIDE	
PHOSPHATASE	PULLULAN PUL	
Glycine ARYLAMIDASE	D-RAFFINOSE	
ORNITHINE DECARBOXYLASE	O/129 RESISTANCE (comp. vibrio.)	
LYSINE DECARBOXYLASE	SALICIN	
DECARBOXYLASE BASE	SACCHAROSE/SUCROSE	
L-HISTIDINE assimilation	D-TREHALOSE	
COUMARATE CMT	ARGININE DIHYDROLASE 2	
BETA-GLUCURONIDASE	OPTOCHIN RESISTANCE	
O/129 RESISTANCE (comp.vibrio.)		
Glu-Gly-Arg-ARYLAMIDASE		
L-MALATE assimilation		
ELLMAN		
L-LACTATE		

Serial dilution for the soft coral and sponge homogenates to 10<sup>-6</sup>, then 0.1 ml were streaked on six general bacterial media (Marine Agar (Oxoid), R2A Agar (Difco Lab), Starch casein Agar (HIMEDIA), ISP2 Medium (Difco Lab), Actinomycetes Medium (HIMEDIA) and M1 Agar (Oxoid) and incubated at 25°C for 7 days, starting from the third day to the seventh day colonies with different characters were picked up and re-streaked again to obtain pure cultures (Wilson *et al.*, 2010).

# Identification of associated bacteria (Moehario et al., 2019).

Identification of bacterial isolates after Gram staining was performed by using VITEK 2 (bioMerieux). It is an automated full system used for bacterial identification. It works by evaluating each biochemical reaction present in microbial identification cards by using optical signals. Suspension with unknown bacteria is inoculated and then incubated with identification cards.

### **RESULTS**

# Collection of Soft oral and sponge:

# **Sponge samples:**

Eight samples collected from 5 sponges species were collected from red sea as shown in Table 2.

Table 2: Sponge classification

Name	Type	Number of Samples
Ircinia strobilina	Sponge	1
Callyspongia viridis	Sponge	1
Suberea spp.	Sponge	2
Spongia officinalis	Sponge	2
Biemna ehrenbergi	Sponge	2
Total		8

### **Soft coral Isolation**

Twelve samples collected from 5 soft corals species were collected and as shown in Table 3.

**Table 3: Soft corals classification** 

Name	Туре	Number of samples
Heteroxenia fuscescens	Soft Coral	2
Didemnum moseleyi	Soft Coral	2
Lobophytum pauciflorum	Soft Coral	4
Sarcophyton trocheliophorum	Soft Coral	3
Sinularia spp.	Soft Coral	1
Total		12

# Isolation of Soft coral and sponge associated bacteria Soft Coral associated bacteria

Thirty two bacterial isolates were isolated and identified from soft coral samples as identified in Table 4

Table 4: Bacterial isolates from soft corals

Soft Coral species	Number of Soft	Bacterial species	No. of Isolates
•	coral samples	•	
Didemnum Moseleyi	2	Aerococcus viridans	1
		Alloiococcus spp	3
		Staphylococcus hominis	1
Heteroxenia Fuscescens	2	Vibrio spp.	2
		Helcococcus kunzii	3
Lobophytum Pauciflorum	4	Vibrio spp.	1
		Staphylococcus epidermidis	5
		Aerococcus viridans	3
		Staphylococcus gallinarum	1
		Staphylococcus saprophyticus	3
Sarcophyton Trocheliophorum	3	Enterococcus faecalis	2
		Granulicatella Elegans	2
		Staphylococcus vitulinus	2
Sinularia spp.	1	Lactococcus garvieae	2
		Arthrobacter creatinolyticus	1
Total	12		32
Percent from the total 54 isolates			59 %

## Sponge associated bacteria

Twenty two bacterial isolates were isolated and identified from Soft Coral samples as identified in Table 5

Table 5: bacterial isolates from Sponges.

Sponge species	Number of sponge samples	Bacterial species	No. of Isolates
Biemna ehrenbergi	2	Kocuria rosea	5
		Kocuria kristinae	2
Callyspongia viridis	1	Kocuria rosea	4
Spongia officinalis	2	Bacillus pumilus	2
		Staphylococcus cohnii	1
		Staphylococcus xylosus	2
Suberea spp.	2	Kocuria rosea	3
Ircinia strobilina	1	Kocuria kristinae	3
Total	8		22
Percent from the total			41 %
54 isolates			

### **DISCUSSION**

Marine environment is considered one of the most diverse environments The Red Sea is one of these global hotspots for (Choudhary et al., 2018). biodiversity contain numerous types of coral and sponges (DiBattista et al., 2016). This study was intended to show the bio diversity for the Marine Macro and Microorganism. Soft corals were classified as; Heteroxenia fuscescens, didemnum moseleyi, Lobophytum pauciflorum, Sarcophyton trocheliophorum and Sinularia spp. which was reported by Ibrahim et al. (2014), Afifi et al. (2016), Hassan et al. (2016), Tarek et al. (2016) and Zubair et al. (2016), . Sponges were classified to: Ircinia strobilina, Callyspongia viridis, Suberea spp, Spongia officinalis and Biemna ehrenbergi, this was reported by Ilan et al. (2004), El-Ganainy et al. (2005), Mohamed et al. (2014) and Shaala and Almohammadi (2017) (2014). In this study bacterial isolated had been isolated from soft coral and sponges as following, Aerococcus viridans, Alloiococcus spp., Enterococcus faecalis, Vibrio spp., Granulicatella elegans, Helcococcus kunzii, Kocuria kristinae, Kocuria rosea, Lactococcus garvieae, Bacillus pumilus, Arthrobacter creatinolyticus, Staphylococcus cohnii, Staphylococcus lentus, Staphylococcus xylosus Staphylococcus epidermidis, Staphylococcus hominis, Staphylococcus saprophyticus and Staphylococcus vitulinus. Generally bacterial isolates from Soft coral was thirty two isolated with 59 % which was higher than sponge which was twenty two isolates with 41 %.

That was hypothesized to that sponge is more aggressive than soft corals which make corals develop defense mechanism by having more types of bacteria to grow (Aerts and Rob, 1997).

## **CONFLICT OF INTEREST**

The present study was performed in absence of any conflict of interest.

### REFERENCES

Aerts, L. A. M. and Rob, V. S. (1997). Quantification of sponge/coral interactions in physically stressed reef community, NE Colombia. Marine Ecology-progress Series - MAR ECOL-PROGR SER., 148: 125-134.

- Afifi, R.; Abdel-Nabi, I. M. and El-Shaikh, K. (2016). Antibacterial activity from soft corals of the Red Sea, Saudi Arabia. J Taibah Univ Sci., 10(6): 887-95.
- Chen, Y. H.; Kuo, J.; Sung, P.J.; Chang, Y. C., Lu, M.C.; Wong, T. Y.; Liu, J. K.; Weng, C. F.; Twan, W. H. and Kuo, F. W. (2012). Isolation of marine bacteria with antimicrobial activities from cultured and field-collected soft corals. World Journal of Microbiology and Biotechnology., 28(12): 3269–3279.
- Choudhary, A.; Rai, D.; Cotter, P. and Dobson A. (2018). Marine antimicrobials for the food industry. Tresearch, (13): 2.
- DiBattista, J. D.; Roberts, M. B. and Bouwmeester, J. (2016). A review of contemporary patterns of endemism for shallow water reef fauna in the Red Sea. J Biogeogr., 43: 423–439.
- El-Ganainy, A.; Yassien, M. and Ibrahim, E. A. (2005). Bottom trawl discards in the Gulf of Suez, Egypt. Egyptian J. Aquat. Res., 31.
- Hassan, M. H. A.; Rabab, M.; Mona, H.; Tarek, M.; El-Gendy, O. A. and Sleim M.
  A. (2016). Biological and Chemical Investigation of the Soft Coral Lobophytum pauciflorum Collected from the Egyptian Red Sea. International Journal of Pharmacognosy and Phytochemical Research., 8: 906-911.
- Ibrahim, S. R.; Mohamed, G. A.; Shaala, L. A.; Youssef, D. T. and Gab-Alla, A. A. (2014). Didemnacerides A and B: Two new glycerides from Red sea ascidian Didemnum species. Natural Prod. Res., 28, 1591-1597.
- Ilan, M.; Gugel, J. and van Soest, R. (2004). Taxonomy, reproduction and ecology of new and known Red Sea sponges. Sarsia., 89: 388–410.
- Kotb, M. M. A.; Hanafy, M. H.; Rirache, H.; Matsumara, S.; Al-Sofyani, A. A.; Ahmed, A. G.; Bawazir, G. and Al-Horani, F. (2008). Status of coral reefs in the Red Sea and Gulf of Aden Region. In: Status of Coral Reefs of the World. 2008, 67-78.
- Loya, Y. (1972). Community structure and species diver- sity of hermatypic corals at Eilat, Red Sea. Marine Biology, 13: 100-123.
- Moehario, H.; Boestami, P.; Edbert, D.; Enty, T. and Robertus, T. (2019). Automation for the identification of Pseudomonas aeruginosa: Comparison of TDR-300B, VITEK®2, and VITEK®-MS. Bioraxi. 10, 1101.
- Mohamed, G. A.; Ibrahim, S. R. M.; Badr, J. M. and Youssef, D. T. A. (2014). Didemnaketals D and E, bioactive terpenoids from a Red Sea ascidian Didemnum species. Tetrahedron., 70: 35–40.
- Rheinheimer, G. (1992). Aquatic Microbiology, John Wiley, New York. 3.
- Rinkevich, B. (2005). What do we know about Eilat (Red Sea) reef degradation? A critical examination of the published literature. Journal of Experimental Marine Biology and Ecology., 327: 183-200.
- Ritchie, K. B.; Schwarz, M; Mueller, J.; Lapacek, V. A.; Merselis, D.; Walsh, C. J. and Luer, C. A. (2017). Survey of Antibiotic-producing Bacteria Associated with the Epidermal Mucus Layers of Rays and Skates. Frontiers in Microbiology, 81050.
- Shaala, L.A. and Almohammadi, A. (2017). Biologically active compounds from the red sea sponge Suberea sp. Pak. J. Pharm. Sci., 30(6): 2389-2392.
- Sheppard, C.; Price, A. and Roberts, C. (1992). Marine ecology of the Arabian region: patterns and processes in extreme tropical environments. Academic Press, London.
- Stoeckl, N.; Farr, M.; Larson, S.; Adams, V.M.; Kubiszewski, I.; Esparon, M. and Costanza, R. (2014). A new approach to the problem of overlapping values: A case study in Australia's Great Barrier Reef', Ecosystem Services. 10, 61-78.

- Tarek, M.; Ali, A. A.; El Komi, M.; El Hadi, M. and Fayez, S. (2016). Growth Rate Assessment of Alcyonacean Sarcophyton glaucum Northern Hurghada, Red Sea, Egypt. Natural Resources, 7: 384-398.
- Wilson, G. S.; Raftos, D. A.; Corrigan, S. L. and Nair, S. V. (2010). Diversity and antimicrobial activities of surface-attached marine bacteria from Sydney. Microbiol. Res., 165(4). 300–311.
- Zubair, M; Alarif, W.; Al-Footy, K.; Mohamed, P.H.; Ali, M. and Basaif, S. (2016). New antimicrobial biscembrane hydrocarbon and cembranoid diterpenes from the soft coral Sarcophyton trocheliophorum. Turk. J. Chem., 40(3): 385-92.