Effect of different concentrations of Ethylenediaminetetraacetic Acid (EDTA) on growth performance and physiological aspects of Nile Tilapia, *Oreochromis niloticus* Fingerlings.

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ABSTRACT

The present study deals with the effect of different concentrations of EDTA (0, 0.1 & 0.3 g EDTA /L) on the growth in length, weight, survival rate, growth performance and physiological characters of the Nile tilapia, *Oreochromis niloticus* fingerlings reared in glass aquaria. The improvement of fish fingerlings development obtained by EDTA is dose-dependent. Fingerlings obtained from high concentration of EDTA (0.3 g EDTA /L) showed higher increase in length, weight and survival rate than those of low concentration (0.1 g EDTA /L) and control. The total weight gain, average daily gain and specific growth rate of *O. niloticus*, fingerlings improved with the high concentration of EDTA compared with the low concentration and control. The best feed conversion ratio (1.82) was recorded in the fish fingerlings exposed to 0.3 g EDTA/L compared with the control (2.1) and fingerlings exposed to 0.1 g EDTA/L (2.44). Biochemical analysis indicated that, EDTA in high concentrations are useful additives for isolated or/ and reducing dangerous substances from different fish tissues and surrounding media, thus improved the physiological responses.

Keywords: EDTA; growth performance; physiological aspects; Oreochromis niloticus.

INTRODUCTION

A variety of chemicals and drugs are used in aquaculture for some purposes such as sediment and water management, enhancement of natural aquatic productivity, transport of live organisms, feed formulation, manipulation and enhancement of reproduction, growth promotion, health management, processing and adding value to the final product (Douet *et al.*, 2009). Those include associated with structural materials, soil and water treatments, antibacterial agents, pesticides, feed additives, anesthetics and hormones (GESAMP, 1997).

EDTA has been used extensively in medicine as a chelating agent for removal of the toxic heavy metals. The disodium salt of EDTA is a common component in many eye drops and contact lens wetting and cleansing solutions. EDTA is also used in a number of personal care and hygiene products, such as shampoos, liquid soaps, creams, and lotions. Household disinfectants often contain EDTA, especially if fatty acid soaps are used in the disinfectant formulation. These soaps are sensitive to calcium and magnesium, and the chelating agent prevents the formation of hard-water soap curds (Hart, 1984). EDTA is a common sequestrant and antioxidant added to foods, body care, and household products. It binds trace minerals such as lead, copper, iron, cadmium and nickel that may be present in the product. When EDTA adds as an antioxidant, it prevents oxygen from causing color changes and rancidity (Ben-Best, 2009). EDTA has two advantages with respect to other compounds–its relative low

biodegradability in groundwater systems (Nowack, 1996) and its strong complexing capacity with heavy metals (Kedziorek and Bourg, 2000).

Tilapia is an ideal candidate for warm water aquaculture. They spawn easily in captivity, use a wide variety of natural foods, as well as formulated feeds, tolerate poor water quality and grow rapidly at warm temperatures. These attributes, along with relatively low input costs have made tilapia widely cultured freshwater fish in tropical and subtropical countries (Biswas *et al.*, 2005; El-Saidy and Gaber, 2005; Fasakin *et al.*, 2005; Borgeson *et al.*, 2006; Tahoun, 2007; Tsadik and Bar, 2007; Al-Absawy, 2010 and Khalaf Allah *et al.*, 2013). In Egypt the Nile tilapia, *Oreochromis niloticus* aquaculture productin about 870938 tons which form 63.48 % from our total aquaculture and natural production and put Egypt as a second bigger productive for Nile tilapia after China. The total production of tilapia fry from hatcheries or fish farms in 2012 attained 242.558 million fry of *O. niloticus and O. aureus* dominate the fry production (GAFRD, 2012 and FAO, 2014).

The effect of chelating agent (EDTA) as benefit in reduce lead and cadmium toxicity with improve the physiological and biochemical profiles of the fish and its impact on growth performance were investigated by few authors (Shalaby, 2007; Shalaby et al., 2011 and Tonsy and Abdel-Rahman, 2012). The previous studies, investigate the improvement of growth performance in the Nile tilapia, Oreochromis niloticus by exogenous treatments, L-thyroxine (T4) hormone (Khalil et al., 2011) and temperature (Hashem et al., 2012). However, there is a little information on the improvement of growth performance and physiological responses on the Nile tilapia by chelating agent, Ethylenediaminetetraacetic acid (EDTA). One of the difficulties in assessing the health state of natural fish population has been the paucity of reliable references in the normal conditions. Physiological and biochemical parameters reflects physical and chemical changes occurring in an organisms, therefore detailed information can be obtained on general metabolism and physiological status of the fish in different groups of age, habitat and different effects of anticoagulants, even in the same species (Kocabatmaz & Ekingen, 1978 and Tavares-Dias & Silva-Sandrim, 1998).

Therefore, the present study aimed to evaluate the effect of different concentrations of Ethylenediaminetetraacetic acid (EDTA) on growth performance and biochemical status of Nile tilapia, *Oreochromis niloticus* fingerlings. Such investigations may lead to a better understanding of some biological and physiological aspects.

MATERIALS AND METHODS

A total of 400 specimens of *Oreochromis niloticus* fingerlings, infected with black spot disease which caused by encysted metacercariae (Fig. 1) were obtained from El-Hadad Special Farm at Kafr El-Sheikh Governorate during October, 2013. Fish transported to Cairo in large plastic bags filled with oxygen, each containing approximately 20 L of fresh water. Duration of the journey was approximately three hours. Fish were acclimatized for one week in well aerated large glass tank; and fed daily on a commercial fish diet.

The experimental work was conducted using 9 separated tanks. Each one (90X55X30 cm) filled with 80 liters fresh dechlorinated water. In each tank, 30 fish were kept randomly selected from the stock tank. Fish were exposed to 0.1 g EDTA /L as treatment group 1, and 0.3 g EDTA /L as treatment group 2 and control. Each group including three tanks.

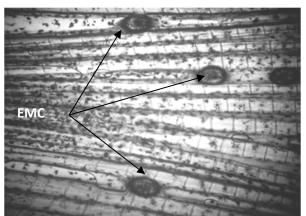


Fig. 1: Photomicrograph of caudal fin clip of *Oreochromis niloticus* fingerlings showing encysted metacercaria (EMC).

All tanks were provided with aeration and thermostastic heater, change of water was carried out and the aquaria were cleaned weekly and add one drop of 1% methylene blue for each tank. Fish in each aquarium were fed twice daily during six days per week by commercial diet containing 35% protein. Fish were fed at a rate of 3 % of fresh body weight. The experiment was conducted for 5 weeks, length and weight of 15 randomly selected fish from each treatment were recorded weekly and their initial weights and survival rate were recorded.

A-Biological Data:

Growth in length, weight and survival rate estimation:

Length and weight of 15 randomly selected fish from each treatment were recorded weekly and their initial weights and survival rate were recorded.

Measurement of growth performance:

Total weight gain, average daily gain, specific growth rate, feed intake (FI) and feed conversion ratio in *O. niloticus* fry were determined according to Recker (1975) and Castell & Tiews (1980) as following:

A-Total weight gain $(mg/fish) = (W_T-W_I)$.

Where: W_T = final means weight of fish in grams.

W_I = initial means weight of fish in grams.

B-Average daily gain (ADG) (g.fish/day) = total gain/ duration period.

C-Specific growth rate (SGR) % / day) = 100×(Ln W_T- Ln W_I)/ duration period.

Where: Ln= Natural log.

D-Feed intake (FI) = fish weight x feeding level /100

E-Feed conversion ratio (FCR) = dry matter intake (g) / total gain (g).

B-Physiological studies:

After dissection of fish, a known weight of the target organ (muscles collected from the fingerlings), are treated with 0.1 and 0.3 of EDTA were kept under freezing condition at 4 C^0 until the biochemical determination.

Tissue preparation:

For determination of the total proteins, total lipids, total carbohydrates and enzymes activities including aspartate aminotransferase (ASAT) and alanine aminotransferase (ALAT), a known weight of each organ was homogenized in saline solution by using the electric homogenizer, for 2 min. The homogenated specimens were centrifuged 4000 r.p.m. for 15 min. at 2 C^0 in a refrigerator centrifuge. The supernatant solution was used directly or stored at 4 C^0 until the latter examinations.

Determination of the total proteins:

Total protein content in the muscles were determined according to the method of Doumas (1975) using a kit of Vitro Scient Company.

Determination of the total lipids:

Total lipids content in muscles were determined according to the method of Kaplan (1984), using a kit of Reactivos GPL Company.

Determination of the total carbohydrates:

Total carbohydrates content in wet muscles were determined according to the method of Singh and Sinha (1977) as follows:

Reagents:

i) Anthrone reagent was prepared by addition of 72 ml sulfuric acid to 28 ml distilled water. While this mixture was still warm, 50 mg of anthrone and 1gm of thiourea were added with a vigorous shaking, then it was used directly after slowly cooling at the room temperature then it stored at 8 C^0 .

ii) Standard solution was prepared by addition of 50 mg glucose to 100 ml distilled water.

• Procedure:

• To determine the total carbohydrate content, 0.1 ml of the aqueous samples was added into a test tube and diluted with 1 ml distilled water then follow with 5 ml freshly prepared anthrone reagent.

• The blank was prepared by adding 5 ml of anthrone reagent to 1.1 ml of the distilled water.

• The standard solution was prepared by adding 0.1 ml of standard solution to 1 ml distilled water and then add 5 ml of the anthrone reagent.

All test tubes of blank, standard and samples were placed in a boiling water-bath for 10 min. Then, it was leaved to cool for 15 min. at the room temperature within a dark place. Reading of absorbance photometrically was recorded for the standard and the samples against blank at wavelength 620 nm.

Calculation:

Total carbohydrates (mg/100ml) = $A_s / A_{st} X n$

Where: A_s = Absorbance of the samples.

 A_{st} = Absorbance of the standard solution.

n= Concentration of standard solution= 0.05 then, results were converted into mg/gm tissue.

Determination of ASAT and **ALAT activities:**

ASAT and ALAT of wet muscles were determined according to the method of Reitman and Frankel (1957) by using a kit of Bioadwic Company.

Statistical analysis:

Results were expressed in tables as mean \pm S.D. Data were analyzed by using analysis of variance (ANOVA) according to Bailey (1981).

RESULTS

I- Effect of EDTA on O. niloticus, fingerlings:

Effect of different concentrations of EDTA on growth in length, growth in weight, survival rate and growth performance of the cichlid fish, *O. niloticus* fingerlings are given in Tables (1-4).

1- Growth and survival rate:

i) Growth in length:

The results in Table (1) show that, *O. niloticus* fingerlings exposed to EDTA (0.1 or 0.3 g/L) improved fingerlings growth. Moreover, treatment with high concentration of EDTA (0.3 g/L) over the rearing period of 35 days gave a slight

increase in length of fingerlings $(3.9\pm0.7\text{cm})$ compared with the control and low concentration which have similar values during the period of study.

Exposure time	Control	Treatments		
(Days)	Control	0.1 g EDTA/L	0.3 g EDTA/L	
0	3.2±0.4	3.2±0.4	3.2±0.4	
7	3.5±0.4	3.5±0.4	3.6±0.4	
14	3.5±0.5	3.5±0.5	3.6±0.5	
21	3.6±0.6	3.6±0.6	3.7±0.6	
28	3.6±0.6	3.6±0.6	3.7±0.6	
35	3.8±0.7	3.8 ± 0.7	3.9±0.7	

Table 1: Increments in standard length (Mean±SD cm) of O. niloticus fingerlings treated with EDTA.

ii) Growth in weight:

Data in Table (2) indicate regular increase in total body weight of *O. niloticus* fingerlings treated with EDTA compared with control. It is obvious that a slight increase is evident at high concentration during the period of experiment. But the reverse was show at low concentration, recorded the lowest value of increments in total body weight compared with control.

Table 2: Increments in total weight (Mean±SD gm), of O. niloticus fingerlings treated with EDTA.

Exposure time	Control	Treatments		
(Days)	Collutor	0.1 g EDTA/L	0.3 g EDTA/L	
0	1.143±0.3	1.143±0.3	1.143±0.3	
7	1.250±0.3	1.248±0.3	1.289±0.3	
14	1.356 ± 0.4	1.352±0.4	1.437±0.4	
21	1.462 ± 0.5	1.457±0.5	1.582±0.5	
28	1.674 ± 0.6	1.641±0.6	1.771±0.6	
35	1.842±0.7	1.768 ± 0.7	1.931±0.7	

iii) Survival rate:

The present results (Table, 3) indicate that, *O. niloticus* fingerlings exposed to EDTA (0.1 or 0.3 g/L) improved survival rate of fingerlings than those of control. The treatment with high concentration of EDTA (0.3 g/L) over the rearing period (35 days) gave higher increase in survival rate of fingerlings (90.9%) declined to 85.1% at 0.1 g/L compared with 60.6% at control. It was also noted that, the external mortality signs of control showing erosion of caudal fin, exophthalmos and abdomen swelling (Figs. 2&3). While, the external mortality signs of treated fish exposed to EDTA showing enlargement in gills associated with open mouth.

Exposure time	Control	Treatments		
(Days)		0.1 g EDTA/L	0.3 g EDTA/L	
0	100	100	100	
7	93.9	97.9	97	
14	81.8	95.7	97	
21	77.3	93.6	97	
28	69.7	91.5	92.4	
35	60.6	85.1	90.9	

Table 3: Survival rate (%) for *O. niloticus* fingerlings treated with EDTA.

iv) Growth performance:

Data in Table (4) show, body weight, total weight gain, average daily gain (ADG g.fish/day), specific growth rate (SGR%/day), feed intake (FI gm/fish) and feed conversion ratio (FCR) for *O. niloticus* treated with EDTA.

The present data show that, the total weight gain of *O. niloticus*, fingerlings improved in general with the increasing concentration of EDTA up to 0.3 g EDTA/L compared with the control and low dose. The highest value of total weight gain (0.788) was detected in the fish exposed to 0.3 g EDTA/L. The same pattern was observed with average daily gain and specific growth rate (Table, 4).

The specific growth rate (S. G. R.) for the fish exposed to 0.3 g EDTA/L (1.50) is higher than the value (1.25) for the fish exposed to 0.1 g EDTA/L compared with the control (1.36). The best feed conversion ratio (1.82) was recorded in the fish exposed to 0.3 g EDTA/L compared with the control (Table, 4).

Item	Control	0.1 g EDTA /L	0.3 g EDTA /L
Initial live weight (g/fish)	1.143 ± 0.3	1.143±0.3	1.143 ± 0.3
Final live weight (g/fish)	1.842±0.7	1.768±0.7	1.931±0.7
Weight gain (g/fish)	0.699	0.625	0.788
Average daily gain (g.day/fish)	0.020	0.018	0.023
Specific growth rate (%day)	1.36	1.25	1.50
Feed intake (FI) (g/fish)	1.468	1.523	1.434
Feed conversion ratio (FCR)	2.1	2.44	1.82

Table 4: Growth performance of O. niloticus, treated with different concentrations of EDTA.

II- Physiological studies:

Biochemical analysis in the muscle of fingerling stage *of O. niloticus* reared in glass aquaria for 35 days are showing in Table (5) and are graphically represented in Figs. (4 -8).

Table 5: Biochemical analysis in the fingerling stage muscles of *Oreochromis niloticus*, exposed to the different concentrations of EDTA.

Concentrations (g/L EDTA)	Total Proteins (mg/g wet wt.)	Total Lipids (mg/g wet wt.)	Total Carbohydrates	ASAT (U/g wet wt.)	ALAT (U/g wet wt.)
			(mg/g wet wt.)		
Control	211.67 ± 7.02	70.00 ± 2.00	7.47 ± 0.28	6.00 ± 0.20	0.90 ± 0.10
0.1 g/L	262.33 ± 12.42	65.17 ± 2.75	4.10 ± 0.35	5.63 ± 0.15	$0.67\pm0.0.6$
0.3 g/L	297.33 ± 8.74	58.97 ± 2.59	3.57 ± 0.51	5.37 ± 0.15	0.53 ± 0.06

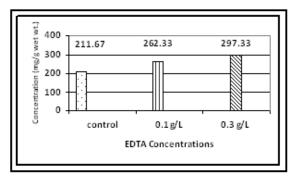


Fig. 4: Changes of total proteins in the muscles of treated *Oreochromis niloticus* fingerling with different concentrations of EDTA.

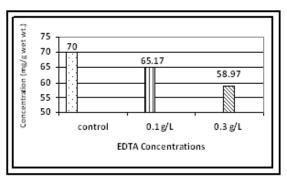


Fig. 5: Changes of total lipids in the muscles of treated *Oreochromis niloticus* fingerling with different concentrations of EDTA.

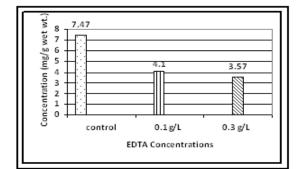


Fig. 6: Changes of total carbohydrates in the muscles of treated *Oreochromis niloticus* fingerling with different concentrations of EDTA.

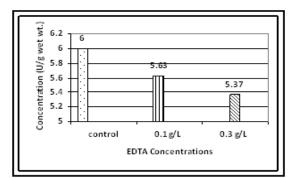


Fig. 7: Changes of ASAT activity in the muscles of treated *Oreochromis niloticus* fingerling with different concentrations of EDTA.

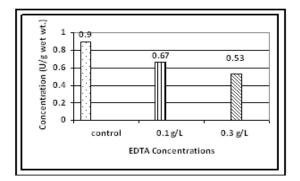


Fig. (8): Changes of ALAT activity in the muscles of treated *Oreochromis niloticus* fingerling with different concentrations of EDTA.

The present results showed that, an addition of EDTA to aquaria contain fingerling stage of *O. niloticus* affects growth and biochemical parameters due to enhancement in metabolic parameters and enzymes activities at different concentrations compared with control samples.

Table (5) and Figure (4) show a significant difference of total proteins in the muscles of *O. niloticus* fingerlings reared in aquaria treated with different concentrations (0.1 g/L and 0.3 g/L) compared with control samples; being 262.33 ± 12.42 mg/g wet wt. in the lowest dose and 297.33 ± 8.74 mg/g wet wt. in the highest one compared with 211.67 ± 7.02 mg/g wet wt. for control. Anova (P<0.05) indicated that, a highly significant differences were obtained at the different concentrations of EDTA.

However, a slightly decreases was observed in the concentration of total lipids in muscles of fingerling stage, *O. niloticus* at low concentration of EDTA treatments (0.1 g/L) and reached its lowest value at high concentration of treatments (0.3 g/L) compared with control; being 65.17 ± 2.75 mg/g wet wt. for minimal dose and 58.97 ± 2.59 mg/g wet wt. for the maximal one compared with 70.00±2.00 mg/g wet wt. for control samples (Table, 5 and Figure, 5). Statistically, (P<0.05) showed highly significant difference at the high concentration of EDTA.

Results are shown in Table (5) and Figure (6) attained a reverse decreasing in total carbohydrates in the muscles of *O. niloticus* fingerlings exposed to EDTA with increasing in dose concentration compared with control. The minimum value of total carbohydrates was recorded in the samples exposed to 0.3 g/L (3.57 ± 0.51 mg/g wet wt.). It showed a slightly increase at the samples treated with 0.1 g/L (4.1 ± 0.35 mg/g

wet wt.) but the highest value was recorded for control group $(7.47\pm0.28 \text{ mg/g} \text{ wet wt.})$. Analysis of variance (P<0.05) exhibited that, a highly significant differences were recorded at different concentrations.

The present study exhibited that, the depletion of aspartate aminotransferase (ASAT) in the muscles of *O. niloticus* fingerlings was observed with increasing the dose of EDTA compared with control; being 5.63 ± 0.15 U/g wet wt. at low concentration (0.1 g/L) and 5.37 ± 0.15 U/g wet wt. at high concentration (0.3 g/L) compared with 6.00 ± 0.20 U/g wet wt. for the control samples (Table, 5 and Figure, 7). Anova (P<0.05) indicate a significant difference at high concentration of EDTA.

Alanine aminotransferase activity (ALAT) in the muscles of fingerling stage of *O. niloticus* exposed to different concentrations of EDTA showed acute depletion with increasing in concentration of the dose compared with control samples; being 0.53 ± 0.06 U/g wet wt. at the high concentration (0.3 g/L) and 0.67 ± 0.06 U/g wet wt. at the low concentration (0.1 g/L) compared with 0.9 ± 0.10 U/g wet wt. in the samples of control (Table, 5 and Figure, 8). Statistical analysis (P<0.05) show a significant differences at the different concentrations.

DISCUSSION

This study aims to evaluate the effect of different concentrations of Ethylenediaminetetraacetic acid on growth parameters, growth performance and survival rate of *O. niloticus* fingerlings. Tilapians are considered as the best species for culture, because of their high tolerance to adverse environmental conditions, ease of reproduction, their fast growth and potential for domestication (EL-Sayed, 1999).

In the present study, the treatment with 0.3 g EDTA /L accelerated fingerlings growth, since a higher increase in growth in length, growth in weight of fingerlings occurred during the experimental period. Treated fish fingerlings gave also a higher survival rate than that of control. Similar reports showed that, EDTA improves survival rates and accelerate growth and development in teleost fish (Shalaby, 2007; Shalaby *et al.*, 2011; El-Banna and Atallah, 2009 and Tonsy & Abdel-Rahman, 2012).

The improvement of fish fingerlings development obtained by EDTA at the present study is dose-dependent. Fingerlings obtained from high concentration of EDTA (0.3 g EDTA /L) showed higher increase in length, weight and survival rate than those of low concentration of EDTA (0.1 g EDTA /L). No abnormalities were observed in fish fingerlings from treated with EDTA. However, the data suggest that a concentration of 0.3 g EDTA /L is appropriate for accelerating growth and improving survival during the experimental period. Similar observations were obtained by Shalaby (2007); Shalaby *et al.* (2011); El-Banna and Atallah (2009) and Tonsy & Abdel-Rahman (2012).

On the other point of view, Kasetsart (2011) studied the effect of different concentrations of EDTA on the development of oyster, *Crassostrea belcheri* and mentioned that, no significant differences in mean percentages of embryos that developed into normal D-larvae were found among untreated and treated units at Na₂-EDTA concentrations ranging from 5 to 20 ppm (P > 0.05). Non-significant differences were found among the mean percentages of fertilized eggs that developed into normal trochophore larvae in untreated seawater and seawater pre-treated with Na₂-EDTA and conditioned from 0 to 48 hr (P > 0.05).

The growth promoters improve the immunity, productivity and economic efficiency of fish via its improvement body weight of the fish (Carnevali *et al.*,

2006), weight gain (Venkat *et al.*, 2004), feed conversion ratio and efficiency (Abdel-Hamid and Mohamed, 2008).

In the present study, the best feed conversion ratio was recorded in fish fingerlings (1.82) when the fish treated by 0.3 g EDTA /L compared to control (2.1). This result was higher than that recorded in the same species in other farms. El-Shebly (1991& 1998) recorded food conversion of 1.8 in Serow Fish-Farm and 1.21 in fresh water fish farm at Manzalah Lake. Chelators such as EDTA have been reported to possess antibacterial activity because of the ability of these chelating agents to disrupt bacterial cell membranes by removing essential divalent cations required to link lipopolysaccharide molecules in the outer membrane (Vaara, 1992). Furthermore, strong chelates may inhibit microbial metabolism by binding trace mineral required for cellular reproduction, growth and survival (Boziaris and Adams, 1999).

Culture assays indicate that, both EDTA and EDTA-chelated trace metals enhance the growth of phytoplankton in sea water (Johnston, 1964). Two mechanisms have been proposed for the enhancement of algal growth by chelating agents such as EDTA. Johnston (1964) suggested that, EDTA increases the solubility and thus the availability of trace metals that are necessary for growth. However, an alternative hypothesis is that chelating agents reduce the availability of harm metals to phytoplankton by lowering the concentration of harm free metal ions (Sunda and Guillard, 1976). In this hypothesis, enhanced algal growth in the presence of EDTA is explained by reduced inhibition by toxic metals which are present in sea water (Davey *et al.*, 1973; Sunda and Guillard, 1976 and Ragan *et al.*, 1980).

The present study revealed that, the average values of final body weight, total weight gain and average daily gain (ADG g/fish/day) increase in the fish fingerlings when the fish treated by 0.3 g EDTA /L compared to control. Results came in close agreement with those obtained by Shalaby (2007); Shalaby *et al.* (2011) and El-Banna & Atallah (2009). Tonsy & Abdel-Rahman (2012) mentioned that, growth performance in *O. niloticus* was improved due to EDTA application and the effects were more pronounced at 1.5% EDTA/kg diet, which are considered as the optimum dose that improve the health status and growth parameters of fish.

The present results showed that, an addition of EDTA to aquaria contains fingerlings of *O. niloticus* affects on growth and biochemical parameters due to enhancement in metabolic parameters and enzymes activities at different concentration compared with control samples.

A significant increase of total proteins was observed in the muscles of *O. niloticus* fingerlings reared in aquaria treated with different concentrations of EDTA (0.1 g/L and 0.3 g/L) compared with control samples. This increasing may be attributed to the additional EDTA to the media isolate the poisonous substances from the different tissues. These findings which agree with those of Morel *et al.* (1987); Kargin (1996); Abdel-Rahman *et al.* (2009); Rengaswamy *et al.* (2009) and Tonsy & Abdel-Rahman (2012). On the other hand, Shalaby (2003) reported a significant decreasing in these values with exposure to Cd only or with low levels of EDTA at 45 days and attributed his findings to great demands and cellular damage that occurred in the tissues of Cd - toxicated fish as Cd toxicity may cause protein breakdown.

However, total lipids in the muscles of fingerling stage, *O. niloticus* exhibited a slightly decrease with increasing concentration of EDTA compared with control. This may be due to EDTA application and its effects was more pronounced at 0.3 g/ L EDTA, which are considered as the optimum dose that improve the healthy status and growth parameters of the fish in this experiment. Similar observations were reported by James & Sampath (1999) and Nicula *et al.* (2011). They mentioned that, all the

tested biochemical parameters and growth performance were improved due to EDTA application and their effects were more evident at 1.5% EDTA/ kg diet. But differ with Shalaby (2007) on *Oreochromis niloticus* and Shalaby *et al.* (2011) on *Clarias gariepinus*, who indicated that, the addition of EDTA lowered total lipids concentration in the fish, exposed to cadmium toxicity to be similar to that of the control fish.

In the present results, a reverse decreasing in total carbohydrate in the muscles of *O. niloticus* fingerlings exposed to EDTA with increasing in the dose concentration were less than that showed in the control samples. This may be attributed to the depletion in tissues glycogen could have resulted because of its utilization for supplying the energy to the fish under stressed conditions. Similar observations were obtained by Dheer *et al.* (1986); Al-Akel *et al.* (1988) and Ghanem (2006) whom confirmed the fact that, all stress conditions invariably lead to retardation of growth and alter the physiological mechanism. If the stressed condition continuous, long enough mortality ensues.

Also, the results exhibited the depletion of aspartate aminotransferase and alanine aminotransferase (ASAT & ALAT) activities in the muscles of *O. niloticus* fingerlings were observed with increasing the dose of EDTA compared with control. The significant changes in activities of different enzymes (ASAT & ALAT) may be due to the tissues impairment caused by stress conditions. Similar observations were obtained by James *et al.* (1991); Svoboda (2001) and Shalaby *et al.* (2011) and differ with Yamawaki *et al.* (1986) and Shalaby (1997) who stated that, the increase of ASAT and ALAT may be attributed to the hepato-cellular damage or cellular degradation by these heavy metals, perhaps in liver, heart or muscle. However, the decrease of ASAT and ALAT activities in the muscles may be attributed to a number of reasons such as leakage from liver and muscles into the blood, actual liver and muscles enzymes inhibition by the effect of toxicant, disturbance in Kreb's cycle and damage of liver and kidney cells that affect the membranes permeability which in turn liberate the enzymes to extra-cellular fluid and blood (Abou El-Ella, 1996; Salah El-Deen *et al.*, 1996& 2000; Zaghloul, 1997; Abbas, 1998 and Ghanem, 2006& 2011).

CONCLUSION

The growth performance of Nile tilapia, *Oreochromis niloticus* fingerlings are good pattern in the fingerlings treated by 0.3 g EDTA /L. These observations suggest that, the exogenous treatment (0.3 g EDTA /L.) may have practical utility in the fish culture and improve the physiological status.

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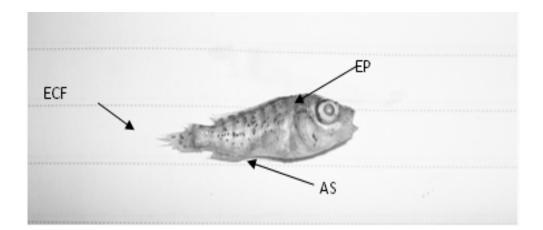


Fig. 2: The external mortality signs of control showing erosion of caudal fin (ECF), exophthalmos (EP) and abdomen swelling (AS).

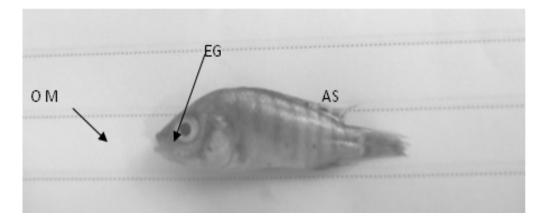


Fig. 3: The external mortality signs of treated fish showing the enlargement in gills (EG) associated with open mouth (OM).

ARABIC SUMMARY

تأثير تركيزات مختلفة من مادة الإيثيلين ثنائى الأمين رباعي حمض الخليك (إيدتا) على النمو، الأداء والخصائص الفسيولوجية لإصبعيات البلطى النيلى "أوريوكرومس نيلوتيكس"

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من أولي المشاكل الخطيرة والهامة في كثير من دول العالم هي نقص البروتين الحيواني اللازم للغذاء وسد الفجوة الغذائية بين الكميات المطلوبة و التي تتزايد عاما بعد عام و الكميات المنتجة من البروتين الحيواني. لذا إتجهت جهود الدولة نحو الإرتقاء بقطاع الإستزراع السمكي.

وحيث أن تحفيز نمو وأداء الأسماك في المزارع السمكية هو الشغل الشاغل لكثير من الباحثين، لذا فإن هذه الدراسة تناولت تأثير التركيزات المختلفة لمادة ايثيلين ثنائي الأمين رباعي حمض الخليك (إيدتا) (١, . جم/ لتر، ٣, . جم/ لتر) على النمو في الطول والوزن ومعدل البقاء وأداء النمو والمظاهر الفسيولوجية لإصبعيات البلطي النيلي المرباة في أحواض زجاجية.

سجلت نتائج الدراسة أن أعلى القيم في معدل زيادة الوزن، الزيادة اليومية في الوزن ومعدل النمو الخاص كانت في مجموعة الأسماك المعرضة للتركيز الأعلى (٣. جم/ لتر) مقارنة بالتركيز الأقل (١. جم/ لتر) والمجموعة الضابطة. كما بينت الدراسة ان أعلي نسبة تحول غذائي (١,٨٢) في مجموعة الأسماك المعرضة للتركيز الأعلى مقارنة بالتركيز الأقل (٢,٤٤) والمجموعة الضابطة (١,١٠).

كما أكدت التحاليل البيوكيميائية أن إضافة مادة الإيثيلين ثنائى الأمين رابع حامض الخليك لها آثار مفيدة فى تحسين الصفات الفسيولوجية مثل البروتين الكلى والدهون الكلية والكربوهيدرات الكلية وكذلك النشاط الإنزيمى لإنزيمى ASAT & ALAT فى عضلات الإصبعيات. كما سجلت النتائج أن التركيز الأعلى من هذه المادة هو الأنسب على الإطلاق لزيادة جودة وصلاحية الأسماك للإستهلاك الآدمى مقارنة بالتركيز الأقل وكذلك المجموعة الضابطة.

وتشير نتائج هذا البحث إلي أفضلية إستخدام مادة الإثيلين ثنائي الأمين رباعي حمض الخليك (إيدتا) عند التركيز ٣. جم/ لتر في تحسين نمو وأداء إصبعيات البلطي النيلي في المزارع السمكية.