The Cunaxid Mite *Neocunaxoides andrei* (Baker & Hoffmann) as a Biological Control Agent of the Root-Knot Nematode *Meloidogyne javanica* Chitwood

Salwa M. E. Shoala^{*} and Gamal A. El Kady^{**}

*Plant Protection Research Institute, Dokki, Giza. **Faculty of Agriculture, Suez Canal University, Ismailia

ABSTRACT

Feeding capacity of the cunaxid mite *Neocunaxoides andrei* (Baker & Hoffmann) and its feeding capability on the rootknot nematode *Meloilogyne javanica* Chitwood, under laboratory or semi field conditions were studied. Results showed that, a female of *N. andrei* consumed an average of 177.2 second juvenile stage (j2s) of *M. javanica* within five days under the laboratory conditions of $26 \pm 1^{\circ}$ C and $70 \pm 5^{\circ}$ R.H. Data also indicated that, the presence of 20, 40 and 60 newly emerged *N. andrei* females together with 500 j2s *M. javanica* in pots planted with tomato seedlings 15 days old caused a reduction of about 59 %, 74 % and 86 % of *M. javanica* reached adult females after 30 days.

Key Words: Cunaxid mite, *Neocunaxoides andrei*, Root-knot nematode, *Meloilogyne javanica*, Biological control, Feeding capacity.

INTRODUCTION

The root-knot nematodes, Meloidogyne species are major yield- limiting pathogens in many crop production areas allover the world. Genus Meloilogyne is considered of a major economic importance (Karssen and Hoenselaar, 1998). In Egypt, the root-knot nematodes are recognized as major agricultural pests of a wide range of crops including, vegetables, fruits and ornamental crops (Oteifa, 1964; Oteifa and Tarjan, 1965). The Meloidgyne species as M. incognita Chitwood, M. javanica Chitwood and M. arenaria Chitwood are widely distributed in northern Egypt (El-Saedy et al., 1993 and Ibrahim et al., 1994). The geographical distribution of the root-knot nematode M. javanica is most abundant in sand soil and newly reclaimed land such as Nubaria, Tahrir province and Salhia districts followed by M. incognita (El-Gindi et al., 1980; Mahrous 1991 and El-Shawadfy, 1997).

Some predacious mites were recorded as a biological control agent of nematodes. Taha *et al.*, (1988) studied the effect of feeding *N. andrei* on the nematode *Panagrolimaus rigidus* Schneider on its developmental time and fecundity under laboratory conditions of 30°C and 70% R.H.. Walter and Kaplan (1991) found that *Coloscerius simplex* (Ewing) colonized greenhouse pot cultures of the root- knot nematodes (*Meloilogyne spp.*) when it fed on vermiform nematode and other soil arthropods. They also studied feeding behavior of Cunaxid mites. Mostafa *et al.*, (1997) reported that *Lasioseius dentatus* Fox could develop on egg masses of *M. javanica* under laboratory conditions.

El-Khateeb (1998) reared *Coloscerius aegyptiacus* Gomaa & El-Khateeb on the free-living

nematode, *Rhabiditella miscicola*, while Sholla (2000) reared *Coloscerius buratus* Den Heyer on the free-living nematode, *R. miscicola* at 26°C and 70% R.H.

El-Hady and El-Naggar (2001) studied the possibility of using both predacious laelapid mites *Hyoaspis bregetovae* (Shereef & Afifi) and *H. sardoa* Berlese as biological control agents of the Root-Knot nematode on sunflower plants. On the other hand, Maareg *et al.,* (2005) reported seven predacious mites species from sugar beet field. These mites were evaluated for their predacious activity on immature stages of *M. incognita.* The results revealed that all tested soil mites except *Cunaxa sp.* fed on immature stage of nematode.

However, the aim of the present work is to study the effect of the predatory mite *Neocunaxoides andrei* (Baker & Hoffmann) as a biological control agent on the root-knot nematode *M. javanica* under laboratory and semi field conditions.

MATERIALS AND METHODS

Rearing cells technique:

Rearing cells made of a closed round transparent plastic containers measuring 5 cm in diameter and 2 cm height filled up to 0.5 cm with plaster of Paris and charcoal of (9:1 w/w) and closed tightly. The rearing cells were maintained at incubator at $26 \pm 1^{\circ}$ Cand $70 \pm 5\%$ R.H.

Preparation of pure culture of the predacious mite *N. andrei*:

Females of *N. andrei* collected from soil samples were transferred singly to rearing cells supplied daily with enough numbers of juvenile stages (j2s) root- Knot nematode M. javanica as a food source. The deposited eggs of the mite were left to develop and the resulted females were mounted for identification.

Root- knot stock culture:

The original inoculum of the root-knot nematode M. javanica was obtained from galled roots of tomato grown in the field of Faculty of Agriculture, Suez Canal University. Nematode egg masses were collected from nematode galls and introduced to tomato roots. Newly hatched juvenile stages were obtained by incubating egg masses on modified Baermann units.

Semi-field condition:

Tomato seeds were sown in small plastic pots (15-cm in diameter) containing 500 g steam sterilized sandy-peat soil (1:1 v/v) under greenhouse condition about 31.5°C ranged from 30°C to 34°C. After 15 days, the tomato seedlings were inoculated with 500 j2s M. javanica/pot around the root. Adult 20, 40 and 60 newly emerged adult cunaxid females were added/pot as soon as the nematode inoculation was conducted. Another pot of tomato plant was inoculated with nematodes only as a control check. The reduction of nematode caused by mites was assessed after 30 days. Water was added carefully when needed. The plants were uprooted and the population of the female nematodes was determined in roots following the method used by Franklin (1949). The experiment was conducted using a complete randomized design with three replicates.

Laboratory condition:

For food consumption of N. andrei on the rootknot nematode M. javanica, about 100 j2s were added daily during five days. The consumed j2s were removed and counted.

Statistical analysis: Data obtained were statistically analyzed by using Costat software programs, Twoway ANOVA for significant differences between means.

RESULTS AND DISCUSSION

Data given in table (1) indicated that the majority of nematode j2s penetrated the tomato roots within 5 days. Mean number of j2s consumed by N. andrei knot nematode M. incognita on sunflower plants by under laboratory condition ranged from 30.8 ± 6.3 to certain predaceous mites. Mites were added to the 35.8 ± 4.16 five days before the nematode injected the soil one to three days before inoculation of roots (Table 2).

Results given in table (3) show that the average number of female nematode decreased significantly

as the number of the cunaxid mite increased. The caused reduction reached 86.2%, 74% and 59.3%, when the cunaxid mites per pot were 60, 40 and 20 females, respectively. The average number of nematode females was 122.6 individuals per control pots.

The average number of galls decreased significantly as the number of the cunaxid mite increased. The caused reduction reached 84.9% %, 70.2 % and 57.4 %, when the cunaxid mites per pot were 60, 40 and 20 females, respectively. The average number of galls was 124.5 individuals per control pots.

The average number of nematodes per 500g soil decreased significantly as the number of the cunaxid mite increased. The reduction reached 81.1%, 80.0% and 76.9% when the cunaxid mites per pot were 60, 40 and 20 females, respectively. The average number of nematodes per 500g soil was 2166.5 ± 53.2 individuals per control pots.

The present results agree with that recorded by Taha et al., (1988) who studied the effect of different prey on the development and fecundity of the predacious mite N. andrei and found that when mites fed on Panagroliaimus rigidus (Schneider) nematode, the immatures developed faster and adult females laid a greater numbers of eggs than those fed on acarid mites.

Also obtained data agree with that found by Mostafa, et al. (1997) who studied the biological control of M. javanica infecting tomato, using the laelapid predaceous mite Lasiosieus dentatus (Fox). Aldicarb treatment and L. dentatus applied either at the same time or 40 days after, showed remarkable improvement on tomato growth.

On the other hand, Amin et al. (1999) found that the highest reduction in total number of the nematodes occurred when the mite L. athiasae was applied four days before nematode inoculation compared with adding mites one day and two days before nematode inoculation.

Also El-Hady and El-Naggar (2001) studied the possibility of the control of egg masses of the rootnematode on roots of sunflower plants. This resulted in reduction of the total number of nematodes. Highest reduction in total nematodes occurred when the mites were applied 3 days before nematode

Average No. Juvenile	Days					Mean of
	1 st	2 nd	3 rd	4 th	5 th	total
Min. No.	420	290	180	40	5	
Max. No.	470	420	230	90	20	
Means \pm S.D	448±19.2	366±50.29	202±19.2	68±19.2	11±5.47	219±17.28

Table (1): Daily average number of a live *Meloilogyne javanica* juvenile injected in 500 gm soil within 5 days under 26 ± 1^{0} C and 70% R.H.

Table (2): Daily average number of consumed *Meloilogyne javanica* by *Neocunaxoides andrei* under laboratory condition $(26 \pm 1^{0}C \text{ and } 70\% \text{ R.H})$ with in 5 days.

No. consumed juvenile M. javanica								
Days	Minimum	Maximum	Average \pm S.D					
1^{st}	22	40	30.8 ± 6.33					
2 nd	30	45	37.8 ± 5.03					
3 rd	24	42	33.8 ± 7.14					
4^{th}	36	48	40.5 ± 4.41					
5^{th}	29	40	35.8 ± 4.16					
Mean			177.2 ± 7.8					

Table (3): Number of the root-knot nematode *Meloilogyne javanica* affected by different rates of the cunaxid mite *Neocunaxoides andrei*.

Doromotor	No. cunaxid mites/pot			Control (free of		
Faranieter	20 mites	40 mites	60 mites	mites)		
Avg. No. nematode females	50 ± 0.8 b	32 ± 0.8 c	$17 \pm 0.5 \text{ d}$	122.6 ±50.1a		
Avg. No. of nematode galls	$53 \pm 6 b$	37.3 ±10.2 c	$18.7 \pm 6.7 \text{ d}$	124.s ±12.4 a		
Avg. No .of nematode female/galls	3 ± 1	2 ± 1	2 ± 1	4 ± 1		
Avg. No.Nematodes/500gm soil	500 ± 20 b	$433. \pm 11.5c$	$408.5 \pm 55.7c$	2166.5 ±53.2 a		
Many fallowed has the same latter do not differ significantly $(n < 0.05)$						

Means followed by the same letter do not differ significantly ($p \le 0.05$).

inoculation.

Maareg et *al.*, (2005) recorded seven predacious mites *Proprioseiopsis messer* (Wainstein), *Cheyletus malaccensis* (Oudemans), *Cunaxa sp., Glycyphagus domesticus* (De Geer), *Macrochelus monchasolska* (B&K), *Platyeseus major* (Halbert) and *Uropoda misella* (Berlese) from sugar beet field. These mites were evaluated for their predacious activity on immature stages of *M. incognita*. The results revealed that the afore-mentioned soil mites except *Cunaxa* sp. fed on immature stages of nematode. On the other hand, Sholla, 2007 reported that numbers of egg masses of *M. javanica* were reduced by adding *N. andrei* to soil planted with cowpea and egg plant.

As a result, it could be concluded that the cunaxid mite *N. andrei* is considered an active biological control agent of the root-knot nematode *M. javanica*.

REFERENCES

Amin, A.W.; Mowafe, M.H. and Ali, Fatma S. 1999. Effect of predaceous mesostigmatid mites in the control of *Meloidogyne javanica* root-knot nematode on kidney bean. Pakistan J. of Nematology, 17 (1): 91-96.

- El-Khateeb, H. M. 1998. Life tables of some predacious mites and their importance in biological control. Ph.D. Thesis, Fac. Agric. Cairo Univ.: 107 109.
- El-Saedy, M. A.; Hassan, M. W. A. and Sourour, M. M. 1993. Occurrence of plant parasitic nematodes and their effects on the nutrition of six fruit crops grown in Nubaria region. J. Agric. Sci., Mansoura Univ., 18: 530-537.
- El-Shawadfy, M. M. 1997. Geographical distribution of root-knot nematodes and their threats to agriculture in Egypt. The 1st Intr. Afro-Asian, Nematology Workshop, 39-44.
- El-Gindi, D. M.; Salem, A. A. and Hashem, M. H. 1980. Structure of plant-parasitic nematode community in weedy soils in Sharquia Governorate effect of soil types. Zagazig J. Agric. Res., 7(1):371-381.
- El-Hady, Mona. M. and El-Naggar, H. I. 2001. Possibility of the control of the root-knot nematode *Meloidogyne incognita* on sunflower plants by certain predaceous mites. J. Agric Sci., Mansoura Univ., 26(4): 2391-2395.
- Franklin, M. T. 1949. A cotton blue lavtophenol technique for mounting plant parasitic nematodes. J. Helminthol, 23: 175-178.
- Ibrahim, I. K. A.; El-Saedy, M. A. and El-Sherbiny, A. A. 1994. Survey study of phytoparasitic nematodes associated with some grasses, weeds

and plants in northern Egypt. J. Agric. Sci., Mansoura Univ., 19: 937-982.

- Karssen, G. and Hoenselaar, T. Van 1998. Revision of the genus *Meloidogyne* Goldi, 1992 (Nematoda: Heleroderidae) in Europe. Nematologica, 44: 713-788.
- Maareg, M. F.; Gohar, I. M. A. and Rady, G. H. 2005. Predatory behavior of some soil mites towards root-knot nematode, *Meloidogyne incognita* infecting sugarbeet crop. Egypt. J. Agric Res., 83 (2): 527-536.
- Mahrous, M.E. 1991. Phytonematodes associated with olive (*Olea europeani*) in newly reclaimed sandy soil with special reference to root-knot nematodes (*Meloidogyne* spp.) Zagazig, J. Agric Res., 18(1): 187-192.
- Mostafa, F. A.; Fouly, A. H. and El-Sherif, A. G. 1997. Biological control of *Meloidogyne javanica* infecting tomato by the predaceous mite *Lasiosieus dentatus*. Egypt. J. of Agronematology, 1(1): 113-120.
- Oteifa, B. A. 1964: A taxonomic guide to the common genera of soil plant nematodes with a supplement on current known economic parasitic species of U.A.R. Contribution of the National

Research Center, pp. 27.

- Oteifa, B. A. and Tarjan, A. C. (1965). Potentially important plant-parasitic nematodes present in established orchards of newly reclaimed sand areas of the U.A.R. Plant Dis Reptr., 49:596-597.
- Sholla, Salwa, M. E. 2000. Taxonomical and biological studies on some predacious Actinedid mites M. Sc.Thesis, Fac., Agric. Ain Shams Univ.
- Sholla, Salwa, M. E. 2007. Studies on some predacous and parasitic mites belonging to suborder Actinedida Ph.D. Thesis, Fac., Agric. Ain Shams Univ., pp. 77.
- Taha, H. A.; El-Naggar, M. E. E., Abou-El-Naga, M. M. and Soliman, S. M. 1988. Effect of different prey species on the development and fecundity of the predacious mite, *Neocunaxoides andrei* (Baker and Hoff.) (Acari: Cunaxidae). Agric., Res., Revi., 66(1): 129-135.
- Walter, D. E. and Kaplan, D. T. 1991. Observations on *Coloscerius simplex* (Acarina : Prostigmata), a predatory mite that colonizes greenhouse cultures of root- knot nematode (*Meloilogyne spp.*) and a review of feeding behavior in the Cunaxidae., Expr. Appl. Acarol., 12 (1-2): 47-59.