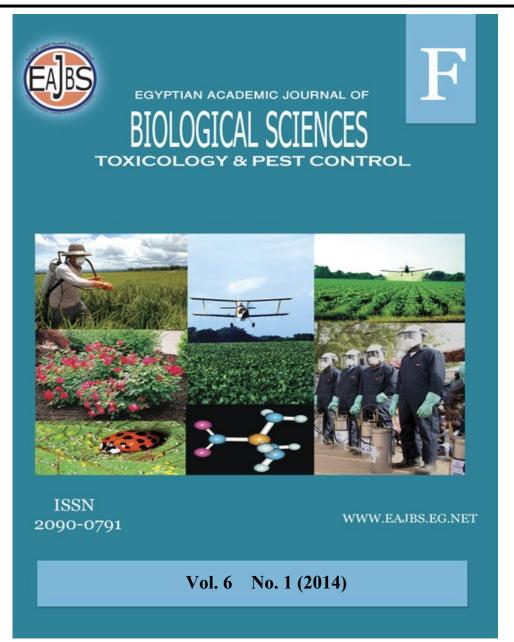
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Impact of some factors on the migration rate and the dispersal of entomopathogenic nematodes

Azazy, A. M *; Saheir F. El-Lakwah and Heba A. A. Al-ghnam Department of Pest Physiology, Plant Protection Research Institute, Agricultural Research Center, Dokki, Giza, Egypt. E-mail: topic68@gmail.com

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ABSTRACT

The migration and the dispersal of five entomopathogenic nematode infective stages had been studied in column of sandy soil under laboratory condition. It was found that it differ according to some factors, these factors included comparison between the effect presence or absence of the insect host Gallaria mellonella and Spodoptera littoralis, insect feces, the nematicide (Nemacur), and host species. The nematicides improved migration and average net distance of all tested nematode strains, except the Steinernema glaseri strain, with which a remarkable inhibition in its mobility, was obviously, observed. Feces of S. littoralis increased the migration rate and the dispersal than the host itself. Most nematode strains were attracted to S. littoralis more than to G. mellonella larvae.

INTRODUCTION

nematodes Entomopathogenic (EPNs) belonging the families to Heterorhabditidae and Steinernematidae are considered excellent biocontrol organisms against numerous insect pests worldwide (Georgis et al., 2006). They are, symbiotically associated with bacteria of genera Photorhabdus and Xenorhabdus, respectively (Akhurst, 1993). The bacteria are carried and maintained by the EPN free-living stage, the infective juvenile (IJ) (Sugar et al., 2012). Once the nematodes locate the insect, they actively penetrate the body cavity and release the bacteria in the hemocoel thereby killing the insect, generally in a short time (Boemare, 2002). The digested insect tissues serve as medium for the nematode and bacterial development. and several generations are produced inside the cadaver. When the resources are depleted and excretion products become limiting, a new cohort of IJs is developed, which acquire bacteria and emerge in search of new hosts (Adams and Nguyen, 2002). The majorities of individuals of *S. carpocapsae* have a sit and wait (ambusher) strategy and tends to be near the soil surface this species was effective against larvae of A. ipsilon, which feeds near or at the soil surface, while H. bacteriophora has an active foraging (cruiser) strategy and occurs deeper in the soil profile.

It was effective against larvae of Otiorhynchus sulcatus, which occurs near roots (Kaya & Gauglar, 1993). These nematodes are faced with a wide array of environmental conditions during the nonfeeding infective stage. Migration and host-finding ability are essential processes in their success as biological control agents. So, the aim of the work to study many factors affecting dispersal and migration of five nematode infective stages (IJs) such as presence or absence of the insect host Gallaria mellonella and Spodoptera littoralis, insect feces, the nematicide (Nemacur), and host species.

MATERIALS AND METHODS Nematodes

Five entomopathogenic nematode species were used in the present study; belong two to the family Heterorhabditidae and three to the family Steinernematidae. The five species were obtained from regular culture in the Laboratory of Insect Parasitic Nematodes, Plant Protection Research Institute, Agriculture Research Centre, Egypt. The entomopathogenic nematode strains, *S. carpocapsae* (All) (S. c. (All)), *S. glaseri* (S. g.), *S. carpocapsae agroitis* (S. c. a), *H. bacteriophora* (HP88) imported from Florida, USA and *H. taysearae* (Ht) (from Giza) by Shamseldean *et al.*, 1996. **The greater wax most, Galleria mellonella.**

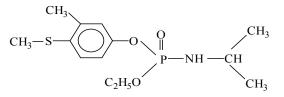
Mass rearing of *G. mellonella* larvae used in the present study was initiated from specimens of bees, heavily infested with the insect which was collected from the apiary of Plant Protection Research Institute, Agricultural Research Center, Dokki, and Giza.

The cotton leafworm, *Spodoptera littoralis*:

Some pupae were obtained from the Department of cotton Insect pests, Plant Protection Research Institute, Agricultural Research Center, Dokki, Giza.

Toxic effect of Fenamiphos (Nemacur) on nematodes:

Fenamiphos (Nemacur) 40% EC, as a nematicide



O-ethyl-O-(3-methyl-4-methyl thiophenyl)-N-isopropyl phosphoroamidate.

HP88 and Ht were used in this study. Six fold serial dilutions of (25, 50,100,200,400 and 800ppm) of Fenamiphos were prepared from the stock solution of the formulated pesticide and distilled water. Ten ml of each chemical dilution was placed in a petri dish. Nematodes were placed into dilutions at a rate of 2000 IJs per dish. Similarly the control contained 2000 IJs but maintained in only distilled water. The treatments were tri-replicated and kept at 25°C. Nematode mortality was calculated once after 48h. and were classified to "not responding to mechanical stimulation" or alive (i.e. moving in response to mechanical stimulation or actively moving). Data was adjusted according Abbott's formula.

Dispersal and migration rate of nematode species:

Such course of investigation was carried out on newly harvested nematode infective juveniles (IJs). According to the foraging strategy of the tested nematode species, the column assay (Azazy, 2001) was used, Plastic tube (20 cm in length and 4 cm in width), was filled with sandy soil (10% w/w) and divided into four equal sections (5 cm length for each) Fig.1.

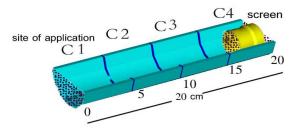


Fig. 1: Dispersal and migration assay unite.

Full grown larvae of G. mellonella or 6th instar larvae of *S. littoralis*, or feces of S. littoralis were used as a bait and kept inside a wire screen cage (1mm hole size) filled with moist sand, and placed at one end of this plastic tube (B). The prepared tubes were incubated at 25±1°C in the dark, for 24h to allow equilibration of any diffuses from the insects and feces through the sand column before applying the nematodes. Six thousand IJs in 3 ml distilled water were, applied to the site of application of each column and incubated at $25\pm1^{\circ}$ C for 24h. After incubation period every sand soil of each section was transferred to a separate petri dish, containing four G. mellomella or S. littoralis larvae and dishes were incubated at the same temperature. Mortality percent was recorded, dead larvae were dissected in the end of the 5th day after infection and numbers of adult nematodes were recorded. Nematode dispersal and migration rate were quantified by:

a) The migration rate "%m"; the percentage of IJs recovered outside the site of application.

b) The average net distance E (D) traveled by the IJs outside the site of application.

E (D) is calculated according to the following formula:

$$\mathbf{E}(\mathbf{D}) = \frac{\sum_{i=1}^{5} n_i x_i}{\mathbf{N}}$$

Where *i* is one of five sand sections; *ni* is the number of IJs recovered in C1+C2+C3+C4+C5; xi is the distance between C1and C2 (in the center) of each section and N is the total number of nematodes recovered in the assay unit. Nematodes found in given sections, were expected to have moved in that section half of the corresponding length indicated in the figure. E (D) can range from 0 to 20 cm.

Statistical analysis:

Statistical analysis of data was carried out using a computer software package "costat", a product of Cohort Software Inc., Berkeley, California, USA. Duncan's multiple range test (Duncan, 1955) was used to differentiate between means.

RESULTS

Some factors affecting dispersal and migration of five nematode infective stages (IJs) had been studied in laboratory in column of sandy soil. These factors included the presence or absence of the insect host, host species, insect feces and the nematicide (Nemacur). First, authors studied the toxic effect of nematicide (Nemacur) against entomopathogenic nematodes. HP88 and Ht were used as an example.

Toxic effect of Fenamiphos (Nemacur) on nematodes:

The toxicity effect of nematicide (Fenamiphos) (Nemacur) on Ht and HP88 infective stages during 48h of exposure were shown in Table (1). Results showed that, the Ht mortality, after 48h exposure to Fenamiphos ranged between 3.3 to 15.7%. Fenamiphos gave mortalities of 4.7, 3.3, 3.8, 8.5, 7.3 and

15.7% for concentrations of (25, 50,100,200,400 and 800 ppm), respectively.

Generally the average mortality was 7.3% of Fenamiphos. Statistical analysis revealed that there were no significant differences between the tested concentrations of pesticide. These pesticide achieved H. bacteriophora (HP88) mortality ranged between 1.1 and 6.8, successively. Statistical analysis proved that Fenamiophos were no significant differences between its concentrations.

 Table 1: Mortality percent of the infective stages of *H. bacteriophora* (HP88) and *H. taysearae* (Ht)

 after 48h exposure to Nematicides Fenamiphos (N.)

Nen	naticides (F	enamiph	nos) (N.)				
Concentration		(HP88)			(Ht)		
25 ppm	1.	1±0.6	b	2	1.7±2.3	b	
50 ppm	3.	7±0.3	ab	(°)	3.3±0.4	b	
100 ppm	3.	8±0.6	ab		3.8±1.9	b	
200 ppm	6.	8±3.5	а	8	3.5±5.2	ab	
400 ppm	5.	4±0.9	ab	7	7.3±2.9	ab	
800 ppm	6.	5±1.7	ab	1	15.7±3.3	а	
*L.S.D		5.04			9.4		
One way ANOVA Complete	ly Random	ized					
Main Effect	df	Fv	value	Р			
Fenamiphos on (HP88)	5	1.6		.22 n	s		
Fenamiphos on (Ht)	5	2.3	3	.10 n	S		
*Duncala multiple vange teet	L						

*Dunca's multiple range test

Values in the same column followed by the same letter are not significant different ($P \le 0.05$)

1- Comparison between the presence and absence of insect host on the dispersal and migration of nematodes.

Twenty-four hours post nematode inoculation, majority of IJs moved away from the point of application in all tested nematode strains. The proportion of migration (%m) was not the same for the different nematode species. In case of host absence (Fig. 2: A & B), results showed that the strain Ht was the best (25.5% m), whereas S.c (All) was the lowest one (9.4m %). However,

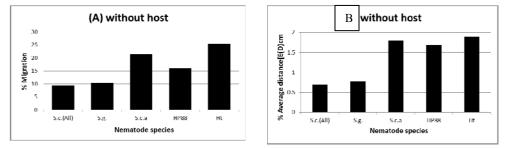


Fig. 2: The migration and dispersal of five nematode strains without hosts

In the case of host presence G. *mellonella* (Fig. 3: A), the migration of IJs of S. g was the highest recording 89.28%. The other strains achieved migration rates of 79.7, 76.48, 29.85 and 21.45% for Ht, HP88, S.c (All) and S.c.a, respectively. Regarding the "average net

distance" [E (D)], in host presence *G. mellonella* (Fig.3: B) IJs of HP88 was found to move an "average net" longer than that of the other strains reaching the maximum distance of 13.07cm. The [E (D)] of the other tested strains were ranged between 3.23 to 12.62 cm.

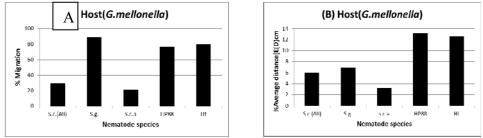


Fig. 3: The migration and dispersal of five nematode strains with Galleria mellonella

The presence of *S. littoralis* as bait increased the migration rates of all the tested strains (Fig.2.A and Fig.4: A). S. g showed the highest values (from 10.43 to 91.12%), followed by HP88 (from 16.1 to 80.06%), Ht (from 25.54 to 34.2%), S.

c. a (from21.5 to 27.3%), S.c(All) (from 9.38 to 22.14). Accordingly, as a result of the increase in % m, an increase in [E (D)] values was evident in all strains (Fig.4: .B).

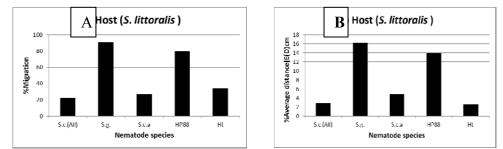


Fig. 4: The migration and dispersal of five nematode strains with host S. littoralis

2- Effect of Host species.

Results showed in (Figs. 3. A and 4. A), revealed that, EPNs varied in their attraction towards the two hosts. Two strains (Ht and S. c. (All)) were attracted more by *G. mellonella* larvae (79.7 and 29.9%m). While; the other three strains (S.g, S.c.a and HP88) were attracted by *S. littoralis* (91.12, 80.6 and 27.3 %m) more than *G. mellonella*.

The variations in the other strains were not too high between the two hosts as in the fore mentioned strains. Regarding E (D) values (Figs. 3: B & 4. B), it is clear that the strain S. g which was attracted by, *S. littoralis* was the superior (16.15cm) followed by the strain HP88 (13.98 cm), which against highest E (D) values (13.1cm) toward *G. mellonella* then Ht was (12.62cm) and the other two strains S.c (All) and S.c.a were the last for the both hosts.

3- Comparsion between the presence host and feces of *S. littoralis:*

As shown in Fig. (5. A), (%m) of all strains (except S. g), in presence of feces of *S. littoralis* were 83.64, 63.89, 52.3 and 42.39% for HP88, S.c.a, S.c (All) and Ht, respectively better than (%m) in presence of the host *S. littoralis* (Fig. 4.A). In contrasts S.g was the least response strain to *S. littoralis* feces

achieving 19.48% of (%m), against 91.12 % migration towards the host itself. In Fig. (5. B), the presence of feces increased values of E (D) of S.c (All), S.c.a and Ht (8.21, 10.14 and 3.96 cm, respectively), but those values were decreased, remarkably in the strains of S.g and HP88 than those of the host.

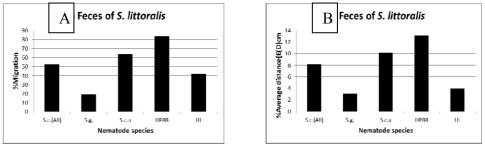


Fig.5: The migration and dispersal of five nematode strains with feces of S. littoralis

4-Effect of Nemacur on the dispersal and migration of nematodes:

Studying the effect of nematicide (Nemacur, 400 ppm) upon the migration of the tested strains (Fig. 6. A), compared with that of *G. mellonella* and *S. littoralis* host upon the migration of the tested strains (Figs.3.A and 4.A), it was found that the nematicide improved the performance of the nematode migration, especially in the case of HP88, Ht, S.c.a,

S.c (All) showing 93.02, 81.13, 68 and 30.86%. On the other hand the nematicide caused clear inhibition in S. g migration (1.08%). Also, strains Ht, S.c.a and HP88 combined with Nemacur, moved longer distances [E (D)] of 10.16,11.9 and 17.15 cm, successively, while those of S.g and S.c (All)were the least, showing 0.1 and 7.6cm, respectively (Fig.6: B).

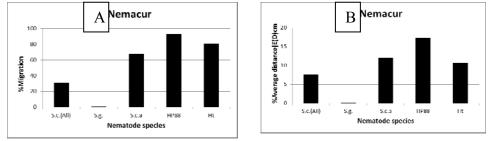


Fig. 6: The migration and dispersal of five nematode strains with Nemacur.

DISCUSSION

Entomopathogenic nematodes of the Heterorhabditidae and Steinernematidae appear to be capable of distance dispersal and local longmigration. Their transmission strategies include both, highly active seek- and behaviors destrov and ambusher strategies, and they may be sensitive to sex related factors in their own populations (Downes & Griffin, 1996). dispersal Although nematode is necessary for host -finding, they may disperse passively (Epsky et al., 1988; Timper et al., 1988), active dispersal,

particularly by cruiser nematodes. appears to be the primary means for host finding, since it increases the likelihood of encountering sedentary host larvae. On the other hand, high mobility without sensitivity to host cues leads to quickly depleted food reserves (Molyneux, 1984 & Vänninen, 1990). Both locomotion and migration ability of the nematodes are thus critical factors in the control of sedentary insect soil. The sand column technique used in the present investigation enabled us to study the dispersal and migration of the tested nematode species, as well as some

factors that may affect them. Also, complying with the situation of sessile root pests in the filed, and host larvae in the assay unit were immobile (Fig.1), allowed the formation of chemical gradient around them (Steiner, 1996). From an ecological perspective, foraging strategies and motility are important variables in population dynamic models (Campos-Herrera *et al.*, 2012). These variables are influenced by abiotic and biotic environmental factors (Lewis *et al.*, 2006) and by intrinsic nematode traits (Campos-Herrera and Gutiérrez, 2014).

In the present investigation, it was found that dispersal and migration differ among the tested strains. These differences could be related to the inherited characters of each species, in addition to the reaction between these features and the other factors. Factors affecting active nematode dispersal and host finding in soil include small pore (Blackshaw spaces and Senthamizheselvan, 1991). moisture, temperature and plant roots (Choo et al., 1989). Finer particle sizes in soil inhibit nematode movement (Noosidum et al., 2010).

dispersal Active of entomopathogenic nematodes is short range and may be influenced by host cues. Host derived compounds such as Carbon dioxide (Co2) (Gaugler et al., 1980) and fecal components (Schmidt and All, 1979) have been shown to be attractive to these nematodes. Migration rate may be directly related to the host finding strategy of tested nematode species. The low migration rate of S. c. (All) and S. c .a. in most experiments typical of an ambusher or a sit and wait strategy (Kaya & Gaugler 1993) and they tended to remain near the point of application limits contact with sedentary hosts (Georgis & Poinar, 1983a; Gaugler et al., 1989). As shown in our result the presence of the host increased the dispersal and migration of the tested nematode species. This finding is in a full

agreement with Georgis & Poinar (1983b) who stated that the presence of the host increased dispersal of H. bacteriophora, but the majority were still found near the placement site. The pattern of dispersal and migration in S.c.a. and S. g. respectively were not affected by changing the host. Also the present results are in accordance with the finding of Grewal et al. (1995) who reported that entomopathogenic nematode species are different in their response to host chemical cues. Understanding the mechanics of foraging behaviors is the key to constructing predictions of how foraging strategy influences nematode biology and the mobile distance that they move. Cruiser species, such as *H. megidis*, directionally respond to host cues and can travel for long distances. Ambushers, however, lack any directional response to host cues and are less mobile. (Chen et al, 2003). S. carpocapsae and S. scapterisci for example, spend most of their time in prolonged bouts of motionless nictation which may last several hours, which is typical of ambushing species. Many Steinernema species exhibit jumping (Campbell and behavior. Gaugler. 1993). The frequency of jumping, like standing behavior, varies among species, and is increased by mechanical contact, air movement, and volatile host cues (Campbell and Kaya, 2000).

The present work proved that the nematicide (Nemacur) is an important factor affecting migration and dispersal of nematode species. This finding agrees with the previous studies, since Ishibashi Shingi, and 1993 stated that the insecticides stimulated the entomopathogenic nematodes to move actively. On other hand (Kamionek 1979) reported that the compatibility between some herbicides and insecticides can diminish host seeking ability and diminish reproductive potential without causing significant mortality in exposed nematode populations. The nematicides

can, probably be used at concentrations of no more than 800 ppm, but insecticides could be used at concentrations more than 1600 ppm with nematodes. Overall, results indicate the feasibility of an IPM use of these nematode species and chemical pesticides in crop protection (Rovesti & Doeso 1990). Further, it is possible that the movement/migration was not only modulated by host attraction but by more selected chemical attractions such as those reported by Choe et al., 2012 and Kaplan et al., 2012. S. feltiae was attracted to both insect and slug associated cues and its strong attraction to slug cadavers suggests that EPNs could also scavenge on carcasses other than those of insects (Nermut et al., 2012).

S. littoralis feces attracted more portions of the tested nematode populations than the host it self and accordingly caused an increase in the average net distances except in the case of *S.g.* strain, since the host attracted the nematode more than feces. This finding confirmed by the work of Schmidt and All (1978, 1979). In the laboratories they found that *S. feltiae* positively responds various stimuli such as Co₂, thermal gradient, and excretory products in insect feces.

Comparing between S. littoralis larvae and G. mellonella larvae in attracting various nematode strains, it was found that G. mellonella attracted about 79.7%m of Ht population while 34.2%m were directed towards S. littoralis larvae. Also the infective stages of the strain S.c (All) preferred the same host with a lees degree. On the other hand the strains S.g. S.c.a and HP88, somewhat preferred larvae of S. littoralis than the other host. Klein-Beekman et al. (1994) found that dispersal of S. g.juveniles was enhanced in the presence of Melolontha melolontha larvae; clear response direction towards the host was not observed. Steiner (1996) reported that unidentified *Steinernema* species and *S. kraussei* exhibited negative migration when using *G. mellonella* larvae as a host. He suggested that *G. mellonella* was repellent to those nematodes. Also, he found that the host finding ability of a strain of *S. feltiae* was smaller for *M. melolontha* than for *G. mellonella*. He attributed this decrease; to that *M. melolontha* feces were repellent to the nematode juveniles.

REFERENCES

- Adams BJ, Nguyen KB. (2002). Taxonomy and systematics. Pp. 1–33 *in* R. Gaugler, ed. Entomopathogenic nematology. Wallingford, UK: CABI Publishing.
- Akhurst, R. J. (1993). Bacterial symbionts of entomopathogenic nematodes, the power behind the thorne. In: Bedding, R& Kaya, H. (Eds.), "Nematodes and biological control of pests". CSIRO, Australia, pp. 127-145.
- Azazy, A. M. (2001). Comparative studies between Insect pathogenic nematodes and other Methods in controlling some soil insects fruit tree borers. Ph.D. Thesis, Fac. Agric., Zagazig University.
- Blackshaw, R.P. and Senthamizhselvan, M. (1991). The effect of sand particle size on *Steinernema feltiae* Filipjev (1934) activity against *Galleria mellonella* larvae. Ann. Appl. Biol. 118: 637-634.
- Boemare N. (2002). Biology, taxomony and systematics of *Xenorhabdus* and *Photorhabdus*. Pp. 35–56 in R. Gaugler, ed. Entomopathogenic nematology. Wallinford, UK: CABI Publishing.
- Campbell, J. F. and Gaugler, R. (1993). Nictation behaviour and its ecological implications in the host search strategies of entomopathogenic nematodes (Heterorhabditidae and

Steinernematidae). *Behaviour* 126, 155-169.

- Campbell, J. F, and Kaya, H. K. (2000). Influence of insect associated cues on the jumping behavior of entomopathogenic nematodes (*Steinernema* spp.). *Behaviour* 137, 591-609.
- Campos-Herrera R. and Gutiérrez C. (2014). *Steinernema feltiae* Intraspecific Variability: Infection Dynamics and Sex-Ratio. J Nematol. 46(1): 35–43
- Campos-Herrera R, Barbercheck M, Hoy WH and Stock SP. (2012). Entomopathogenic nematodes as a model system for advancing the frontiers of ecology. Journal of Nematology 44:162–176.
- Chen, S., HAN, J. LI2, X. and MOENS,
 M. (2003). Effect of temperature on the pathogenicity of entomopathogenic nematodes (*Steinernema* and *Heterorhabditis* spp.) to *Delia radicum*. *Biocontrol* 48, 713–724
- Choe A, Chuman T, von Reuss SH, Dosseyb AT, Yimc JJ, Ajredini R, Kolawa AA, Kaplan F, Alborn HT, Teal PEA, Schroeder FC, Sternberg PW, Edison AS. (2012). Sex-specific mating pheromones in the nematode *Panagrellus redivivus*. Proceedings of the National Academy of Sciences of USA. 109:20949–20954.
- Choo,H.Y.; Kaya,H.K.; Burlando,T. & Gaugler, R. (1989). Influence of plant roots on nematode host finding ability. Environ. Entomol. 18: 1136-1140.
- Downes, M.J. and Griffin,C.T.(1996). Dispersal behaviour and transmission strategies of the entomopathogenic nematodes *Heterorhabditis* and *Steinernema*. Biocont. Sci and Tech., 6: 347-356.
- Duncan, D. B.(1955).Multiple range and multiple F-tests. Biometrics, 11: 1-45.

- Epsky, N.D.; Walter, D.F. and Capinera, J.L. (1988). Efficiency of the *Galleria* (wax moth) baiting technique for recovering infective stages of entompathogenic rhabditis (Steinernematidae and Heterorhabditidae) from sand and soil. Rev. Nemato., 14: 381-387.
- Gaugler, R.; Campbell, J.F. and Mcguire, T.R. (1989). Selection for host finding in *Steinernema feltiae*. J. Invert. Patholo., 54: 363-372.
- Gaugler, R.; Le Beck, L.; Nakgaki, B.and Bouch, G.M. (1980). Orientation of the entomogenous nematode, *Neoaplectana carpocapsae* to carbon dioxide. Environ. Entomol., 9: 649-652.
- Georigs, R. and Poinar, G, O. J. R. (1983b). Effect of soil texture on the distribution and infectivity of *Heterorhabditis bacteriophora* (Nematoda: Heterorabditidae) in sandy loam soil.J.Nematol.15: 652-654.
- Georigs, R. and Poinar, G, O. J.R. (1983a). Effect of soil texture on the distribution and infectivity of *Neoaplectana carpocapsae* (Nematoda: Steinernematidae). J. Nematol., 15: 308-311.
- Georgis R, Koppenhöfer AM, Lacey LA, Bélair G, Duncan LW, Grewal PS, Samish M, Tan L, Torr P, van Tol RWHM.(2006).Successes and failures in the use of parasitic nematodes for pest control. Biological Control., 38:103–123.
- Grewal, P.S; Gaugler, R. and Georgis, R.(1995). Predictors of forging in entomopathogenic strategy nematodes. In: Griffin, G.T.: Gwynn, R. L. & Masson, J.P. (Eds.), "Ecology and transmission strategies of entomopathogenic nematodes. pp: 95-104. European Commission Directorate-XIII, Science, Research and Development, Environ. Research Programme. Official Publications of

the European Communities, 1955, Luxembourg.

- Ishibashi, N. and Shing, T. (1993). Effects of insecticides on movement, nictation, and infectivity of *Steinernema carpocapsae*. J. Nematol., 25 (2):204-213.
- (1979). Influence of Kamionek, M. pesticides on the mortality & effectiveness of N.carpocapsaeWeiser.pp.87-88.In J. proceedings Weiser, of the International Colloquium on Pathol., Inverter. Septemb.11-17, Prague, Czechoslovakia.
- Kaplan F, Alborn HT, von Reuss SH, Ajredini R, Ali JG, Akyazi F, Stelinski LL, Edison AS, Schroeder FC, Teal PE. (2012). Interspecific nematode signals regulate dispersal behavior. PLoS One 7:e38735.
- Kaya, H.K. and Gaugler, R. (1993). Entomopathogenic nematodes. Ann. Rev. entomol., 38: 181-206.
- Klein-Beekman, M. Z.; Wiegers, G. L. and Smits, P. H. (1994). Biological cockcahafer control of larvae (Melolontha melolontha) with the entomopathogenic nematode Steinernema glaseri. Med. Fac. Landbouww. Univ. Gent., 59 (2 a): 411-419.
- Lewis EE, Campbell J, Griffin C, Kaya HK, Peters A. (2006). Behavioral ecology of entomopathogenic nematodes. Biological Control., 38:66–79.
- Molyneux, A.S.(1984). The influence of temperature on the infectivity of heterorhabditid and steinernematid nematodes for larvae of the sheep blowfly, *Lucilia cuprina*. Proc.4<u>th</u> Aust. Appl. Entomol. Res. Conf., Australia: 344-351.
- Nermut J., Vladimir P. Z. A, Zdene K. M. (2012). The response of Phasmarhabditis hermaphrodita (Nematoda: Rhabditidae) and Steinernema feltiae (Nematoda: Steinernematidae) to different host-

associated cues Biological Control, 61: 201-206.

- NoosidumA., Hodson A. K., Lewis E. E., Chandrapatya A. (2010). Characterization of New Entomopathogenic Nematodes from Thailand:
- Foraging Behavior and Virulence to the Greater Wax Moth, Galleria mellonella L. (Lepidoptera: Pyralidae) Journal of Nematology, 42(4): 281–291.
- Rovesti, L. and Deseo, K.V. (1990). Compatibility of chemical pesticides with the entomopathogenic Steinernema nematodes, carpocapsae Wesier and Filipjev Steinernema fltiae Steinernematidae). (Nematoda: Nematologica, 36: 237-245.
- Schmidt, J. and All, J.N. (1979). Attraction of *Neoaplectana carpocapsae* to common excretory products of insects.Enviro.Entomol.8: 55-61.
- Schmidt,J. and All, J.N. (1978). Chemical attraction of *Neoaplectana carpocapsae* (Nematoda: Steinernematidae) to common excretory products of insects. Enviro. Entomol. 7: 605-607.
- Shamseldean, M.M., Abou EL-Sooud,
 A. B., Abd-Elgawad, M.M. and
 Saleh, M.M. (1996). Identification of
 a new heterorhabditid species from
 Egypt, *Heterorhabditis taysearae*, n.
 sp.(Rhabditroa: Heterorhabdrndae).
 Egyptian Journal of Biological Pest
 Control 6, 129-138
- Steiner, W.A. (1996). Dispersal and host finding ability of entomopathogenic nematodes at low temperatures. Nematologica., 42: 243-261.
- Sugar DR, Murfin KE, Chaston JM, Andersen AW, Richards GR, deLéon L, Baum JA, Clinton WP, Forst S, Goldman BS, Krasomil-Osterfeld KC, Slater S, Stock SP, Goodrich-Blair H. (2012). Phenotypic variation and host interactions of *Xenorhabdus*

bovienii SS-2004, the entomopathogenic symbiont of *Steinernema jollieti* nematode. Environmental Microbiology., 14:924–939.

Timper, P.; kaya, H. k. and Gaugler, R. (1988). Dispersal of the entomopathogenous nematode *Steinernema feltiae* (Rhabditida: Steinernematidae) by infected adult insects. Environ. Entomol., 17:546-550.

Vanninen,I. (1990). Depletion of endogenous lipid reserves in Steinernema feltiae and Heterorhabditis bacteriophora on infectivity. Proc. Int. Collog. Invertebr. Pathol. Microb. Control, 5: 232.

ARABIC SUMMERY

تأثير بعض العوامل على معدل الهجرة و الأنتشار للنيماتودا الممرضة للحشرات

احمد محمد عزازى _ سهير فيصل اللقوة _ هبه عبد الجليل على الغنام قسم فسيولوجيا الافات – معهد بحوث وقايه النباتات- مركز البحوث الزراعيه – الدقى – الجيزة

تم اختبار معدل الهجرة والأنتشار للأطوار المعدية لبعض أنواع النيماتودا في حالة وجود العائل أو عدم وجوده باستخدام عمود التربة الذي يبلغ طوله ٢٠سم في وجود يرقات دودةالشمع ودودة ورق القطن كطعم حشري،وذلك في التربة الرملية(١٠%رطوبة) ،وقد وجدت هناك اختلافات متنوعة في قدرة الأفراد المعدية لكل نوع و كانت السلالة S. glaseri الأعلى في معدل الهجرة (٢٨, ٨٩%) قاطعة مسافة مقدار ها ٦.٩ سم خلال زمن قدره ٢٤ ساعة وذلك عند استخدام دودة الشمع كطعم، أما عند استخدام دودة ورق القطن كطعم أعلى معدل للهجرة لها ١٠١٢%وتم قطع مسافة ١٠٩٦سم خلال

و في حالة عدم وجود عائل فقد كانت السلالة Ht الأعلى في معدل الهجرة (٢٥.٥٤%) و قطعت مسافة (١.٩٢ سم) خلال نفس الزمن.

وبقياس القدرة على الانتشار والهجرة وجد أن السلالات المختبرة كانت أفضل في حالة وجود المخلفات البرازية لدودة ورق القطن عن وجود العائل كطعم حشري وقد كانت النتائج كما هي : ٣،٤٢.٣٩، ٢،٤٢.٣٩ البرازية لدودة ورق القطن عن وجود العائل كطعم حشري وقد كانت النتائج كما هي : ۳،٤٢.٣٩ معدل الأنتشار للأنواع (HP88, S.c.a, , S.c(All), Ht) على الترتيب و معدل الأنتشار للأنواع , S.c.a ,Ht , هو (٢.٤،٣٠٤

وكذلك وجد أن قدرة النيماتودا على الانتشار والهجرة كانت أفضل في حالة استخدام دودة ورق القطن كطعم حشري عن استخدام دودة الشمع حيث كانت ٩١.١٣ ، ٣٤.٢، ٨٠.٠٦ % للأنواع S.g ، HP88 ، Htو المسافة التي قطعتها كانت هي S.c.a ، HP88 ، S.g للأنواع S.c.a ، HP88 ، S.g على الترتيب.